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INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 4:		11) International Publication Number: WO 88/ 05823
C12N 15/00, A61K 39/04 G01N 33/569	A2	43) International Publication Date: 11 August 1988 (11.08.88)
. (21) International Application Number: PCT/US (22) International Filing Date: 1 February 1988 (-	Brook, Smith & Reynolds, Two Militia Drive, Lex-
(31) Priority Application Number:	010,0	(81) Designated States: AT (European patent), AU, BE (European patent), CH (European patent), DE (Euro-
(32) Priority Date: 2 February 1987 ((02.02.8	pean patent), FR (European patent), GB (European patent), IT (European patent), JP, LU (European patent)
(33) Priority Country:	1	tent), ŃL (European patent), ŚE (European patent).
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(54) Title: MYCOBACTERIUM TUBERCULOSIS GENES ENCODING PROTEIN ANTIGENS

(57) Abstract

Mycobacterium tuberculosis genes encoding five immunologically relevant proteins have been isolated by systematically screening a lambda gtl1 recombinant DNA expression library with a collection of murine monoclonal antibodies directed against protein antigens of this pathogen. One of the M. tuberculosis antigens, a 65kD protein, has been shown to have determinants common to M. tuberculosis and M. leprae. In addition, genes encoding proteins of other mycobacteria (M. africanum, M. smegmatis, M. bovis BCG and M. avium) have been isolated. Isolation and characterization of genes encoding major protein antigens of M. tuberculosis make it possible to develop reagents useful in the diagnosis, prevention and treatment of tuberculosis. They can be used, for example, in the development of skin tests, serodiagnostic tests and vaccines specific for tuberculosis.

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MYCOBACTERIUM TUBERCULOSIS GENES AND ENCODING PROTEIN ANTIGENS

Description

Background

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Tuberculosis was the major cause of infectious mortality in Europe and the United States in the 19th and early 20th centuries. Dubos, R. and J. Dubos, The White Plague: Tuberculosis, Man and Society, Little Brown & Co., Boston, MA, (1952).

Today, it remains a significant global health problem.

For example, in the United States there are over 20,000 new cases of tuberculosis diagnosed annually. In addition, the steadily declining incidence of tuberculosis evident in preceding years appears to have changed course, reaching a plateau in 1985 and showing an increase in the first half of 1986. Centers for Disease Control, Morbidity/Mortality, Weekly Report, 34:774 (1986); and Centers for Disease Control, Morbidity/Mortality, Weekly Report, 35:774 (1986).

Worldwide, tuberculosis remains widespread and constitutes a health problem of major proportions, particularly in developing countries. The World Health Organization estimates that there are ten million new cases of active tuberculosis per year and an annual mortality of approximately three

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million. Joint International Union Against Tuberculosis and World Health Organization Study Group, <u>Tubercle</u>, <u>63</u>:157-169 (1982).

Tuberculosis is caused by Mycobacterium (M.) 05 tuberculosis or Mycobacterium (M.) bovis, which are the 'tubercle bacilli' of the family Mycobacteriaceae. M. bovis is a species which causes tuberculosis in cattle and is transmissible to humans and other animals, in whom it causes tuberculosis. 10 present, nearly all tuberculosis is caused by respiratory infection with M. tuberculosis. Infection may be asymptomatic in some, but in other individuals, it produces pulmonary lesions which lead to severe debilitation or death. Resistance to tuberculosis is provided by cell-mediated immune 15 mechanisms.

Mycobacteria are aerobic, acid-fast, non-sporeforming, non-motile bacili with high lipid contents
and slow generation times. M. leprae is the etiologic agent of leprosy and, among the other mycobacteria, the only major pathogen. Bloom, B.R. and
T. Godal, Review of Infectious Diseases, 5:765-780
(1983). However, other mycobacterial species are
capable of causing disease. Wallace, R.J. et.al.,
Review of Infectious Diseases, 5:657-679 (1984).
M.avium, for example, causes tuberculosis in fowl
and in other birds. Members of the M.
Avium-intracellularae complex have become important
pathogens among individuals with acquired immunodeficiency syndrome (AIDS). Certain groups of

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individuals with AIDS have a markedly increased incidence of tuberculosis as well. Pitchenik, A.E. et. al., Annals of Internal Medicine, 101:641-645 (1984).

05 Diagnostic and immunoprophylatic measures for mycobacterial diseases have changed little in the past half century. Tuberculin, developed by Koch as a cure for tuberculosis in the late 1800s, is an M. tuberculosis filtrate of complex and poorly-defined composition. It is used as a skin test antigen to detect prior exposure to the bacillus. Enrichment of the protein fraction of this material in the 1930's produced the purified protein derivative (PPD) which is still used to diagnose exposure to tuberculosis. Seibert, F.M. et.al., American Review of Tuberculosis, 30(Suppl.):705-778 (1934). usefulness is limited, however, by its lack of specificity and its inability to distinguish active disease from prior sensitization by contact with M. tuberculosis or cross-sensitization to other mycobacteria. Young, R.A. and R.W. Davis, Proceedings of the National Academy of Sciences, USA, 80:194-1198 (1983).

Bacille Calmette Guerin (BCG), an avirulent strain of M. bovis, has been used widely as a live vaccine against tuberculosis for over 50 years. Calmette, A., C. et.al., Bulletin of the Academy of

Medicine Paris, 91:787-796 (1924). During that time, numerous studies have shown that BCG has protective efficacy against tuberculosis. studies are reviewed by F. Luelmo in American Review 05 of Respiratory Diseases, 125(pt. 2):70-72 (1982). However, more recently, a major trial of BCG in India indicated that such a vaccine was not protective against tuberculosis in this setting. World Health Organization WHO Technical Report Series, 651 10 (1980). Presently available approaches to diagnosing, preventing and treating tuberculosis are limited in their effectiveness and must be improved if a solution is to be found for the important public health problem tuberculosis represents 15 worldwide.

Summary of the Invention

The present invention is based on the isolation of genes encoding immunogenic protein antigens of the tubercle bacillus <u>Mycobacterium tuberculosis</u> (<u>M. tuberculosis</u>). Genes encoding such protein antigens have been isolated from a recombinant DNA expression library of <u>M. tuberculosis</u> DNA. Genes encoding proteins of four additional mycobacteria have also been isolated and restriction maps produced.

In particular, genes encoding five immunodominant protein antigens of the tuberculosis bacillus (i.e., those M. tuberculosis proteins of molecular weight 12,000 daltons (12kD), 14kD, 19kD, 65kD and 71kD have been isolated by probing a lambda gtll expression library of M. tuberculosis DNA with

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monoclonal antibodies directed against M. tuberculosis-specific antigens.

Recombinant DNA clones producing the specific antigenic determinants recognized by the monoclonal antigens were also isolated in this manner. DNA from such recombinant lambda gt11 clones was mapped with restriction endonucleases; the restriction maps for genes encoding the five immunodominant protein antigens (i.e., genes encoding the 12kD, 14kD, 19kD, 65kD and 71kD proteins) were deduced. The nucleotide sequence of three of the genes have been determined and, in each case, the amino acid sequence of the encoded protein has been deduced.

Brief Description of the Drawings

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Figure 1 shows restriction maps of M. tuberculosis DNA. Recombinant DNA clones isolated with monoclonal antibodies directed against the 12kD, 14kD, 19kD, 65kD and 71kD protein antigens were mapped with restriction endonucleases. The insert DNA endpoints are designated left (L) or right (R) in relation to lac Z transcripts which traverse the insert from right to left. Restriction sites are represented as follows: A, Sal I; B, BamHI; E, EcoRI; G, Bg1II; K, KpnI; P, PvuI; S, SacI; X, XhoI.

Figure 2 shows arrays of antigens from M.

tuberculosis recombinant DNA clones probed with rabbit hyperimmune serum. The code of the recombinant DNA clones shown on the numbers filter is: 1, Y3275; 2, Y3274; 3, Y3279; 4, Y3277; 5, Y3247; 6, Y3272; 7, Y3150; 8, Y3254; 9, Y3147; 10, Y3163; 11,

Y3179; 12, Y3191; 13, Y3252; 14, Y3178; 15, Y3180; 16, Y3143; 17, lambda gt11. Clones 1, 5, 6, 7, 9 and 16 are M. tuberculosis recombinants described in the following section. Clones 10, 11, 14 and 15 are 05 M. leprae recombinants expressing epitopes of the 18kD, 28kD, 36kD and 65kD antigens, respectively. Clones 2, 3, 4, 8, 12, 13 are uncharacterized recombinants from the lambda gt11 M. tuberculosis and M. leprae libraries. Clone 17 is a non-10 recombinant lambda gt11 control.

Figure 3 shows arrays of recombinant mycobacterial antigens probed with monoclonal antibodies to assess the extent of cross-reactivity between recombinant protein antigen of M. tuberculosis and of M. leprae. The array of clones is identical to that shown in Figure 2. Antibody probes and the antigen sizes recognized are: 1/, IT-11 (71kD); 2, IT-31 (65kD); 3, IT-16 (19kD); 4, IT-1 (14kD); 5, IT-3 (12kD).

20 Figure 4 shows restriction maps of DNA encoding four proteins (71kD, 65kD, 19kD and 14kD) of M. tuberculosis and four proteins (71kD, 65kD, 19kD and 14kD) of M. bovis BCG. Restriction sites are represented as follows: A, AatII; B, BamHl; C, BclI; D, DraIII; E=EcoRI; G, BglII; H, HinfI; K, 25 KpnI; P, PstI; S, SalI; V, PvuI and X, XhoI.

Figure 5 is a comparison of restriction maps of the gene encoding the 65kD protein of 6 mycobacteria (M. leprae, M. tuberculosis, M. africanum, M. bovis

30 BCG, M. smegmatis, M. avium). Restriction sites are

as follows: B, BamHl; K, KpnI; N, SacI; P, PvuI; S, SalI; X, XhoI.

Figure 6 is the nucleotide sequence of the region containing the <u>M. tuberculosis</u> 19kD gene. The deduced amino acid sequence of the encoded protein is also represented (protein start position, nucleotide 1110; protein stop position, nucleotide 1586).

Figure 7 is the nucleotide sequence of the region containing the <u>M. tuberculosis</u> 71kD gene and the deduced amino acid sequence of the encoded protein.

Figure 8 is the nucleotide sequence of the region containing the M. tuberculosis 65kD gene. The deduced amino acid sequences of the two long open reading frames are presented in one letter code over (540) or under (517) the appropriate triplets.

Detailed Description of the Invention

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The invention described herein is based on the isolation of genes encoding immunogenic protein antigens of the bacillus M. tuberculosis, which is the major etiologic agent of tuberculosis. In particular, it is based on the isolation, using monoclonal antibodies directed against M. tuberculosis—specific antigens, of genes encoding five immunogenic protein antigens of the tuberculosis bacillus; these five antigens are immunodominant. Immunogenic antigens are those which elicit a response from the immune system. Immunodominant protein antigens are immunogenic

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antigens against which the immune system directs a significant portion of its response. Genes encoding M. tuberculosis antigens of molecular weight 12,000 daltons (12kD), 14kD, 19kD, 65kD and 71kD were isolated in this manner.

Isolation and characterization of major protein antigens of <u>M. tuberculosis</u>, as described herein, make it possible to develop more effective tools for the prevention, diagnosis, and treatment of tuberculosis. Identification and isolation of genes encoding five immunodominant <u>M. tuberculosis</u> protein antigens, as well as of the five protein antigens, are described below; uses of the genes and encoded products are also described.

M. bovis BCG DNA clones were also isolated for the genes encoding the 71kD, 65kD, 19kD and 14kD proteins. In order to compare M. bovis BCG and M. tuberculosis genes encoding proteins of similar molecular weight, restriction endonuclease maps were determined for DNA segments containing each of the genes. Restriction maps for each of these genes is represented in Figure 4.

In addition, DNA clones were isolated for the genes encoding the 65kD protein from M. africanum, M. smegmatis and M. avium. Restriction endonuclease maps were determined for DNA segments containing each of these genes. The restriction maps for these genes, as well as for the genes encoding the 65kD protein of M. tuberculosis, M. bovis BCG and M. leprae, are represented in Figure 5.

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I. Construction of a recombinant expression library of M. tuberculosis DNA

A recombinant DNA expression library of M. tuberculosis DNA was constructed using lambda gtll. The library was constructed with M. tuberculosis genomic DNA fragments in such a way that all protein-coding sequences would be represented and expressed. Young, R.A., B.R. Bloom, C.M. Grosskinsky, J. Ivanyi, D. Thomas and R.W. Davis, Proceedings of the National Academy of Sciences, USA, 82:2583-2587 (1985).

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Lambda qt11 is a bacteriophage vector which is capable of driving the expression of foreign insert DNA with E. coli transcription and translation signals. Lambda gtll expresses the insert DNA as a fusion protein connected to the E. coli Betagalactosidase polypeptide. This approach ensures that the foreign DNA sequence will be efficiently transcribed and translated in E. coli. proach is also useful in addressing the problem of the highly unstable nature of most foreign proteins; fusion proteins are often more resistant to proteolytic degradation than is the foreign polypeptide alone. Lambda gt11 and the E. coli strain used (Y1090) have been described previously. Young, R.A. et al., Proceedings of the National Academy of Sciences, USA, 80:1194-1198 (1983); Young, R.A. and R.W. Davis, Science, 222:778-782 (1983). teachings of these publications are incorporated herein by reference. The library constructed in this manner has a titer of 1x 10¹⁰ pfu/ml. and

contains approximately 40% recombinants with an average insert size of 4kB.

II. Screening of the lambda gtl1 M. tuberculosis library with antibody probes

Murine monoclonal antibodies to protein antigens of M. tuberculosis were used individually to
probe the M. tuberculosis recombinant DNA library.
This work is described below and with specific
reference to the 65kD antigen in the Exemplification. The antibodies used as probes and the sizes
of the antigens to which they bind are shown below.

		M. tuberculosis
-	Antibody	Antigen
	IT-3	12kD
15	IT-20	14kD
	IT-19	19kD
	IT-27	19kD
	IT-17	23kD
	IT-29	23kD
20	IT-15	38kD
	IT-21	38kD
	IT-23	38kD
	IT-13	65kD
	IT-31	65kD
25	IT-33	65kD
	IT-11	71kD

Engers, H.D. et al., <u>Infectious Immunology</u>, 51:718-720 (1986).

All monoclonal antibodies were used at approximately 1:200 to 1:300 dilution in 50mM Tris-HCl pH8/150 mM NaCl/.05% Tween 20.

Screening of the lambda gtll recombinant DNA library was performed as described by Young et al. in Proceedings of the National Academy of Sciences, USA, 82:2583-2587 (1985), the teachings of which are incorporated herein by reference. One modification was made in the method described by Young and co-workers: 1% bovine serum albumin was used in place of 20% fetal calf serum to decrease back-ground.

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Briefly, cloned lambda gt11 recombinants were arrayed on lawns of E. coli Y1090. The phage were grown, antigen expression was induced and the antigens were blotted and probed with serum. Detection of signal-producing plaques was performed with a biotinylated secondary antibody system (Vectastain, Vector Laboratories, Burlingame, CA) or with an alkaline phosphatase conjugated secondary antibody system (Protoblot, Promega Biotec, Madison, WI), both used according to manufacturer's instructions. Signal-producing clones were isolated using antibodies directed against protein antigens of molecular weight 12kD, 14kD, 19kD and 65kD and 71kD. In each case, similar numbers of clones were isolated in screens of approximately 105 recombinant plaques. DNA clones encoding the 23kD and 38kD antigens could not be detected with these antibodies, possibly because the native epitope is modified or topographically complex, or because the

antigen-antibody interaction is too weak to be recognized by current detection methods.

III. Probing of Arrays of lambda gtll DNA Clones with Antibody Probes

05 0.2 ml of a saturated culture of Y1090 was added to 2.5 ml of molten LB soft agar, poured onto 100 mm plates containing 1.5% LB agar and allowed to harden at room temperature for 10 min. 100 ul of phage plate stock containing approximately 1011 10 pfu/ml of the lambda gt11 DNA clones of interest were placed into alternate wells of 96-well tissue culture plates. A multi-pronged transfer device was placed briefly in the wells containing phage and then touched lightly to the surface of the plate 15 onto which the soft agar had been poured. plates were then incubated at 42°C for approximately 3 hours, at which point clear plaques approximately 5mm in diameter were visible. The plates were then overlayed with nitrocellulose filters saturated with 20 10mM isopropylthiogalactoside (IPTG) and incubated at 37°C for 3.5 hours. Subsequent processing of filters for detection of antigen was identical to

Immunoscreening of the lambda gt11 library to isolate clones reactive with monoclonal antibodies specific for the 65kD antigen is described in the Exemplification.

the procedures described for screening of lambda

gtll library with antibody probes.

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IV. Recombinant DNA Manipulation

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DNA from recombinant lambda gt11 clones was isolated and mapped with restriction endonucleases by standard techniques. Davis, R.W. et al.,

Advanced Bacterial Genetics: A Manual for Genetic Engineering, Cold Spring Harbor (1980).

Figure 1 shows the genomic DNA restriction map deduced for each of the genes encoding the five M. tuberculosis antigens and illustrates how each of the cloned DNAs aligns with that map. All clones isolated with monoclonal antibodies directed against any single antigen align with a single genomic DNA segment. This indicates that all clones were isolated because they express the protein of interest rather than an unrelated polypeptide containing a similar or identical epitope. In addition, this result suggests that each antigen is the product of a single gene.

The orientation of each DNA insert in the recombinant clones was determined by restriction analysis. Only among the clones for the 65kD antigen were the inserts found in both possible orientations relative to the direction of Lac Z transcription in lambda gtll. This suggests that this protein can be expressed in E. coli from signals independent of those provided by Lac Z. Similar results have been obtained for recombinant DNA clones encoding the 65kD antigens of M. bovis and M. leprae. Thole, J.E.R. et al., Infectious Immunology, 50:800-806 (1985); Young, R.A. et al., Nature, 316:450-452 (1985).

The nucleotide sequences of three regions of the <u>M. tuberculosis</u> DNA were determined: 1) the region containing the <u>M. tuberculosis</u> 19kD gene; 2) the region containing the <u>M. tuberculosis</u> 71kD gene; and 3) the region containing the 65kD gene. The three sequences are represented in Figures 6-8. Sequences were determined using standard techniques, which are described in the Exemplification.

V. <u>Filter hybridization of Insert DNA</u>

10 Arrays of lambda gt11 clones were created as described above and incubated at 42° for 5 hours. The plates were then overlayed with nitrocellulose filters and placed at 4°C for 1 hour. Probe DNA was labelled with ³²P by nick translation. Filter hybridization was performed as described by Davis et 15 al. in Advanced Bacterial Genetics: A Manual for Genetic Engineering, Cold Spring Harbor (1980), the teachings of which are incorporated herein by reference. Hybridization conditions were as follows: 50% 20 v/v formamide, 5x SSPE (1x SSPE is 0.18M NaCl, 10mM $Na_{1.5}H_{1.5}PO_4$, 1mM Na_2 EDTA, pH 7.0), 1x Denhardt's solution (0.02% w/v Ficoll, 0.02% w/v polyvinylpyrrolidone, 0.02% w/v bovine serum albumin), 0.3% NaDodSO₄ at 42°C for approximately 16 hours, fol-25 lowed by washing in 2x SSPE, 0.2% NaDodSO, at 45°C.

VI. Recombinant Antigens Recognized by Rabbit Serum

The response of a second animal to an antigen preparation of \underline{M} . tuberculosis was assessed by

examining the reactivity of rabbit anti-M. tuberculosis hyperimmune sera with recombinant antigens. Cloned lambda gtl1 recombinants were arrayed on lawns of E. coli and probed with the rabbit hyperimmune serum. Anti-M. tuberculosis hyperimmune serum, produced by repeated immunization of rabbits with M. tuberculosis H37Rv culture filtrate, was provided by J. Bennedsen (Statens Seruminstitut, Copenhagen, Denmark). These sera were used at 1:100 dilution.

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These sera produced positive signals with lambda gtl1 clones encoding each of the five M. tuberculosis epitopes which had been isolated with murine monoclonal antibodies (Figure 2). Particularly strong signals were observed with the 65kD and 71kD antigens (Figure 2). These results demonstrate that mice and rabbits can mount an antibody response to the same protein antigens of M. tuberculosis.

Clones for the five M. tuberculosis antigens were detected at similar frequencies in the lambda gt11 recombinant DNA library. Thus, the number and type of antigen-producing clones isolated with polyclonal serum antibodies should reflect the relative titer and deversity of the individual antibodies in this serum.

To determine whether any of the 5 M. tuberculosis antigens are relatively immunodominant in the rabbit humoral immune response to M. tuberculosis, the M. tuberculosis lambda gtl1 recombinant DNA library was screened with the rabbit serum. Forty signal-producing clones were isolated, arrayed on

lawns of E. coli Y1090 and probed with monoclonal antibodies directed against each of the 5 recombinant M. tuberculosis protein antigens. Remarkably, 17 of the 40 clones (43%) reacted strongly with at 05 least one of the four anti-65kD monoclonal antibodies tested. An additional six clones (15%) reacted strongly with the anti-17kD monoclonal antibody (IT-11). This indicates that a large proportion of the anti-M. tuberculosis antibody 10 present in the rabbit serum was directed against the 65kD antigen of M. tuberculosis and suggests that it is a dominant antigen for the rabbit humoral immune response. Seventeen of the clones did not react with any of the monoclonal antibodies tested, 15 suggesting that the rabbit sera may identify M. tuberculosis proteins not recognized by the murine antibodies.

VII. Antigenic Relatedness of M. tuberculosis and M. leprae Proteins

There is evidence that M. tuberculosis and M.

leprae share immunologically important antigens. To
assess this further, an investigation of the exact
nature of the immunological relatedness among
recombinant protein antigens of M. tuberculosis and
M. leprae was conducted.

For each of five \underline{M} . <u>tuberculosis</u> and four \underline{M} . <u>leprae</u> protein antigens, a single recombinant DNA clone containing most or all of the gene of interest was used to express antigen in the following manner. The recombinant phage clones were arrayed on a lawn

of E. coli Y1090, which was then grown and induced for antigen expression.

Antigen immobilized on nitrocellulose filters was then probed with 26 individual anti-M. tuberculosis and M. leprae monoclonal antibodies. Figure 3 05 shows the array of DNA clones used and the results obtained with the anti-M tuberculosis antibodies IT-1, IT-3, IT-11, IT-16, and IT-31, which recognized proteins of 14kD, 12kD, 71kD, 19kD and 65kD respectively. Table 1 details the full results of 10 these cross-screening experiments, showing the reactivity of antigen expressed from individual recombinant DNA clones with each of the individual monoclonal antibodies. Clones were scored as positive only if the signal produced was clearly 15 greater than the background signal produced by the non-recombinant lambda gtll clone included in/each array.

TABLE 1

Reactivity of Monoclonal Antibodies with

Recombinant Protein Antigens

								DNA	CLONES				
					អ. <u>tu</u>	berculo	sis				H. 16	prae	
1	נאג	IBODI	<u>Es</u>	12kD Y3275	14kD Y3247	19kD Y3147	65kD Y3150	71kD Y3272	- 19t11	18kD Y3179	28kD Y3163	36kD Y3180	65kD Y3178
	н.	tuber	culosis										
		12kD	IT-3	①	-	-	-	-	-	-	-	-	-
		14kD	IT-1	-	•	-	-	-	,=		-	-	
			IT-4	-	\odot	-	-	-	-	-	-	_	-
			IT-20	-	\odot	-	-	-	-	-	-	-	-
		19kD	IT-10	•	••	\oplus	-	-	-	-	-	-	-
			IT-12	-	-	\odot	-	-	-	-	-	-	-
			IT-16	-	-	\odot	-	-	-	-	-	-	-
	-		IT-19	-	-	\odot	-	-	•	_	- 9	-	-
		65kD	IT-13	-	-	-	①	-	_	_	- -	-	_
			IT-31	-	-	-	⊕	-		-	_	_	①
			IT-33	-	-	-	⊕	-	-	-	-	-	⊙
		71kD	IT-11	-	-	-	-	•	-	-	-	-	-
	н.	<u>lepra</u>	<u>e</u>	•									
		18kD	L7-15	-	•	-	-	-	-	•	-	-	-
		28kD	SA1.D2D	-	-	-	-	-	-	-	•	.	-
			SAL.BIII	· -	-	-	-		-	-	-	-	-
		3 6kD	F47-9-1	-	-	-	_	-	-	-	-	①	-
			HLO4-A		-	-	-	-	-	-	-	-	-
		65kD	ci.i	-	- -	-	•	-	-	-	-	-	⊙
			IIH9 .	-	-	-	•	-	-	-	-	-	\odot
			IIIE9	-	-	-	. •	-	-	-	•	-	
			IICS	-	-	-	\odot	•	-	-	-	-	$\check{\odot}$
			IIIC8	-	-	-	-	-	-	-	-	-	\odot
			T2.3	-		•	\odot	-	-	-	-	-	•
			Y1-2	-	-	-	①	-	-	-	-	-	$\odot \odot \odot \odot \odot \odot$
			SA2.D7C	-	•	-	①	-	-	-	-	-	⊙
•			HL30A	\odot	①	•	①	①	•	\odot	\odot	\odot	•

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Several conclusions can be drawn from the results shown in Table 1. Among the 11 monoclonal antibodies that recognize a 65kD antigen, 7 react with the 65kD protein from both mycobacteria (IT-31, C1.1, IIH9 (identical to IT-33), IIC8, T2.3, Y1-2, 05 SA2.D7C), one antibody reacts only with the M. tuberculosis 65kD protein (IT-13), and two antibodies react only with the M. leprae 65kD protein (IIIE9 and IIIC8). One antibody, ML30A, cross-reacts with an antigen in E. coli and could not specifically identify antigen-producing clones. These results indicate that the 65kD protein antigens of M. tuberculosis and M. leprae are homologues and share a number of epitopes. addition to these shared epitopes, however, both 65kD antigens have epitopes that are specific for one species relative to the other.

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No cross-reactivity was observed between other antigens of these two mycobacterial species. Because monoclonal antibodies recognize a single 20 epitope and because only one or a few antibodies were available for each antigen, it is not clear whether the 65kD proteins are the only homologous protein antigens of M. tuberculosis and M. leprae. 25 Among the antigens for which lambda gt11 clones have been isolated, the 18kD antigen of M. leprae and the 19kD antigen of M. tuberculosis are of similar size. To determine whether these two antigens are related, the homology of the DNA sequences that encode these antigens was examined. At conditions of moderate 30 stringency, no hybridization was observed between

the insert DNA and Y3147 (an \underline{M} . <u>tuberculosis</u> 19kD clone) and Y3179 (an \underline{M} . <u>leprae</u> 18kD clone). This indicates no significant homology between the DNA sequences of the insert DNAs of these two clones. This result suggests that the \underline{M} . <u>tuberculosis</u> 19kD and the \underline{M} . <u>leprae</u> 18kD proteins are unlikely to be homologous.

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As a result of the work described, recombinant DNA clones encoding five major protein antigens of M. tuberculosis were isolated through the use of an extensive collection of well-characterized murine monoclonal antibodies. These five proteins were also found to be major antigens in the rabbit humoral immune response to M. tuberculosis. One of these antigens, the 65kD protein, is shared with another other mycobacterial pathogen M. leprae.

Several lines of evidence indicate that the 65kD antigen is among the most immunodominant of the protein antigens of M. tuberculosis. Eleven of the 25 different M. tuberculosis and M. leprae monoclonal antibodies examined in this study recognized the 65kD recombinant antigen from one or both mycobacteria. In addition, almost half of the recombinant DNA clones isolated with rabbit polyclonal anti-M. tuberculosis sera express the 65kD antigen, reflecting the predominance of antibody to this antigen in these sera.

Considerable evidence indicates that the 65kD antigen plays an important role in the human response to tuberculosis. Antibodies directed against this protein can be detected in the serum of

patients with tuberculosis. The 65kD antigen is present in purified protein derivatives (PPD's) of M. tuberculosis, M. bovis, and other mycobacteria. Thole, J.E.R. et al., Infection Immunity, 50:800-806 (1985). Finally, helper T cell clones reactive with recombinant 65kD antigen have been isolated from patients with tuberculosis, indicating that this antigen is involved in the cell-mediated as well as the humoral immune response to tuberculosis.

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10 Among the major antigens of the leprosy bacillus, the 65kD antigen appears to elicit antibody and T cell responses similar to those observed for the M. tuberculosis antigen. Both serum antibodies and T cells directed against the 65kD M. leprae antigen have been observed in patients with 15 leprosy. Britton, W.J. et al., Journal of Immunology, 135:4171-4177 (1985); Mustafa, A.S. et al., Nature, 319:63-66 (1986). In addition, T cell clones from leprosy patients have been found to respond to recombinant 65kD protein of M. bovis, as 20 well as to PPD's from both M. bovis BCG and M. leprae. Emmrich, F. et al., Journal of Experimental Medicine, 163:1024-1029 (1986); Shankar, P. et al., Journal of Immunology, 136:4255-4263 (1986). interesting to note that in vaccine trials in Asia 25 and Africa, BCG provided significant protection against leprosy, ranging from 20% to 80%. Fine, P., Tubercle, 65:137-153 (1984). An intriguing possibility is that the M. bovis BCG 65kD antigen is involved in engendering the immune protection 30

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provided by this vaccine against \underline{M} . \underline{leprae} , as well as against \underline{M} . $\underline{tuberculosis}$.

In addition to the 65kD antigen, there is evidence that the 19kD and 71kD antigens of M.

105 tuberculosis may be particularly important in the immune response to this bacillus. Helper T cell clones from tuberculosis patients have been isolated which respond to the recombinant 19kD protein. The 71kD antigen is recognized by the humoral immune system of both mice and rabbits, and antibody to this antigen has been shown to be a prominent component of hyperimmune anti-M. tuberculosis rabbit sera.

VIII. Isolation of DNA Clones for Genes Encoding Proteins of Additional Mycobacteria

Using the procedures described above for isolation of genes encoding M. tuberculosis proteins, genes encoding proteins of additional mycobacteria were isolated. DNA clones containing genes encoding the following proteins were isolated:

	Mycobacterium	Protein	Clone
	M. bovis BCG	71kD	PL1-101
	·	65kD	PL1-105
		19kD	PL1-501
25		14kD	PL1-502
	M. smegmatis	65kD	PL1-206
	M. avium	65kD	PL1-401
	M. africanum	65kD	PL1-301

-23-

For purposes of comparison, genes encoding the following proteins were isolated for <u>M. tuberculosis</u> and <u>M. leprae</u>:

	Mycobacterium	Protein	Clone	
05	M. tuberculosis	71kD	Y3272	
		65kD	Y3150	
		19kD	Y3147	
		14kD	Y3248	
	M. leprae	65kD		

The following strains were used for this purpose:

	Species	<u>Isolate</u>				
	M. leprae	Armadillo isolate (WHO)				
	M. tuberculosis	Erdmann strain				
15	M. africanum	African clinical isolate				
	M. bovis BCG	Danish vaccine strain				
	M. smegmatis	MC^2-6				
	M. avium	AIDS patient isolate				

DNA from recombinant lambda gtll clones was isolated, as described above, and mapped with restriction endonucleases, using standard techniques. Davis, R.W. et al., Advanced Bacterial Genetics: A Manual for Genetic Engineering, Cold Spring Harbor (1980).

Figure 4 presents a comparison of the restriction maps for four genes of <u>M. tuberculosis</u> with the restriction maps for four genes of <u>M. bovis</u> BCG which encode proteins of the same molecular weight. As is evident from the figure, in each case, the

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restriction sites on the two genes (e.g., those on the <u>M. tuberculosis</u> gene and those on the <u>M. bovis</u> gene which encodes a protein of the same molecular weight) are essentially identical. This indicates that the sequence of the genes of the two mycobacteria (at least those encoding these four proteins) are very similar and, therefore, the proteins they encode are also very similar.

Figure 5 presents a comparison of the restriction map for the gene encoding the 65kD protein for the six mycobacteria. As is evident, the restriction maps for the genes encoding the 65kD protein of M. tuberculosis, M. africanum, M. bovis BCG, M. smegmatis and M. avium are essentially identical. The fact that there is no detectable difference among these mycobacteria at the level of the restriction map is an indication that, at least at

this level, the encoded proteins are the same.

As is also evident, the map of the <u>M. leprae</u> 65kD gene has several identical restriction sties in common with those of the other mycobacteria; it also has two sites not found in the other genes and lacks three sites present in the others. This indicates that, at the level of the restriction map, there are similarities in the DNA (and the encoded protein). In addition, however, there are differences apparent at this level.

IX. Diagnostic, Therapeutic and Preventive Applications

The isolation of genes encoding major protein antigens of M. tuberculosis makes it possible to

-25-

address problems which presently exist in diagnosing treating and preventing tuberculosis. Isolation of genes encoding proteins of other mycobacteria, such as <u>M. bovis BCG</u>, <u>M. africanum</u>, <u>M. smegmatis</u> and <u>M. avium</u> makes it possible to address similar problems in diseases which they cause.

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The nucleotide sequence of three of the five genes has been determined. The sequence of the remaining genes can be determined using well-known methods, such as that of Sanger et al. Sanger, F. et.al., Proceedings of the National Academy of Sciences, USA, 74:5463-5467 (1977). The amino acid sequence of each of the immunodominant proteins has been deduced from the nucleotide sequence of the three genes and can be done for the others.

Identification and characterization of the genes for major tuberculosis protein antigens and of the proteins themselves make it possible to develop improved reagents for diagnosis and immuno-prophylaxis of tuberculosis. Proteins antigens encoded by an entire gene, or amino acid sequences (e.g., peptides, protein fragments) which make up the antigenic determinant of a M. tuberculosis antigen (i.e., M. tuberculosis-specific antigenic determinants) may be used in serodiagnostic tests and skin tests. Such antigens would be highly specific to the tuberculosis bacillus and the tests in which they are used would also be highly specific. Highly specific serological tests would be of great value in screening populations for

individuals producing antibodies to M. tuberculosisspecific antigenic determinants; in monitoring the
development of active disease in individuals and in
assessing the efficacy of treatment. As a result,
early diagnosis of tuberculosis will be feasible,
thus making it possible to institute treatment at an
early stage of the disease and, in turn, to reduce
the likelihood it will be transmitted.

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As a result of the work described, it is also possible to determine which segment(s) of the M. tuberculosis antigen is recognized by M. tuberculosis-specific T cells. A mixture of peptides recognized by helper T cells can serve as a specific skin test antigen useful in assessing immunological status (delayed hypersensitivity) of infected individuals and those with whom they come in contact. This specific skin test antigen would be useful in evaluating rapidly the immunological efficacy of anti-tuberculosis vaccines.

It is reasonable to expect that the products encoded by M. tuberculosis genes, particularly those shown to be recognized by helper T cells, are themselves immunogenic and thus useful components of vaccines against tuberculosis. These products include proteins and portions of such proteins (e.g., polypeptides and peptides). For example, one approach to vaccine development is the introduction of genes encoding products (e.g., polypeptides) which provide immunological protection into viruses such as vaccinia virus, or bacteria, such as cultivatable mycobacteria, thus producing a vaccine

capable of engendering long-lasting and very specific immunity. The genes encoding five immunodominant protein antigens of the tuberculosis bacillus, described herein, are useful for that purpose; genes encoding the 65kD, 19kD and 71kD antigens, or a portion thereof, are particularly valuable in vaccine construction.

Because of the similarities in the DNA encoding similarly-sized proteins and, thus, of the encodied proteins themselves, it is possible that, for example, a vaccine effective against two or more of the mycobacteria can be produced.

EXEMPLIFICATION

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Isolation and Analysis of Recombinants Expressing the 65kD M. tuberculosis Antigen

The recombinant DNA library of M. tuberculosis genomic DNA fragments in the lambda gt11 vector was constructed as described above. Recombinant phage lambda RY3143 and lambda RY3146 were used. Young, R.A. et al., Proceedings of the National Academy of Sciences, USA, 82:2583-2587 (1985). Subclones of the mycobacterial DNA inserts in these recombinant phage were constructed in pUC19 or M13mp9 vectors using standard recombinant DNA techniques. Messing, J. and J. Viera, Gene, 19:269-276 (1982). Maniatis, T. et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1982).

Monoclonal antibodies specific for the 65kD antigen were obtained from the Immunology of Tuberculosis Scientific Working Group under a grant from the WHO/World Bank/UNDP Special Program for 05 Vaccine Development. These antibodies included IT-13 (WTB-78), IT-31 (SA2D5H4), and IT-33 (MLIIH9). Coates, A.R.M. et al., Lancet, 2:167-169 (1981). Gillis, T.P. and T.M. Buchanon, Immunology, 37:172-178 (1982). Anti-B-galactosidase antibodies 10 were purchased from CooperBiomedical. Polyclonal rabbit antisera directed against a sonicate of M. tuberculosis strain H37Rv were elicited as described by Minden and co-workers. Minden, P. et al., Infect. Immun., 46:519-525 (1984). Results are 15 shown in Table 2.

-29-

TABLE 2: PATTERNS OF ANTIBODY REACTIVITIES

	Number of Clones	Reactivi	Reactivity with Antibodi			
		<u>IT-13</u>	<u>IT-31</u>	<u>IT-33</u>		
	27	+	+	+		
05	1	+	+	+		
	2	+	_	+		
	3	-	+	+		
	1	+	•			
	2	-	+	-		
10	2	_	_	+		

a: Recombinant clones expressing antigens reactive with the 65kD antigen specific monoclonal antibodies IT-13, IT-31, and IT-33 were isolated as described For the initial screen, a pool of the three antibodies was used; it contained a 1:1000 dilution 15 of each antibody to screen a total of about 8 \times 10⁵ recombinant phage from the lambda gt11-M. tuberculosis library. To determine which monoclonal antibody reacted with which of the 38 plaquepurified recombinants, about 100 pfu of each 20 recombinant phage were inoculated in small spots on a lawn of Y1090. The phage were allowed to grow and induced to synthesize the foreign proteins as described previously. The filters were then reacted with a 1:1000 dilution of one of the monoclonal 25 hybridoma antibodies as described above.

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The lambda gt11-M. tuberculosis library was screened with the monoclonal antibodies specific for the 65kD antigen and clones reactive with them were isolated essentially as described by Young et al. Young, R.A. et al., Proceedings of the National Academy of Sciences, USA, 82:2583-2587 (1985). Briefly, for each 150mm LB plate, 0.6ml of a fresh overnight culture of Y1090 was infected with 1-2 x10⁵ plaque forming units of the library. 3.5-4 hours of growth at 42°C, the plagues were overlaid with a dry nitrocellulose filter which had been saturated with 10mM isopropyl-B-D-thiogalactopyranoside (IPTG). The plates were incubated an additional 3.5-4 hours at 37°C and then removed to room temperature and the position of the filters The filters were washed briefly in TBST (50 mM Tris-HCl, pH 8, 150mM NaCl, 0.05% Tween 20) and then incubated in TBST + 20% fetal calf serum. After 30 minutes at room temperature, the filters were transferred to TBST plus antibody. For the initial screen, the antibody mix contained a 1:1000 dilution of IT-13, IT-31, and IT-33. The filters were incubated with the antibody solution overnight at 4°C with gentle agitation, washed in TBST and reacted with biotinylated goat anti-mouse immunoglobulin, the Vectastain ABC reagent, and developer as described by the manufacturer (Vector Laboratories). After the color had developed the filters were washed with several changes of water and air dried. Phage corresponding to positive signals were twice plaque purified. To determine

which monoclonal antibodies reacted with which of the recombinant phage, about 100 pfu of a purified phage stock were inoculated in a small spot on a lawn of Y1090 bacteria on an LB plate. The phage were allowed to grow and induced to synthesize the foreign proteins as described above. The filters were then reacted with a 1:1000 dilution of one of the monoclonal antibodies. The subsequent steps were the same as for the initial screen.

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10 Western blot assays were carried out as Cells containing phage or plasmids in follows: which the expression of the foreign sequences was under the control of the E. coli lac gene regulatory sequences were induced to synthesize the foreign 15 proteins by incubating the cells in the presence of 2.5mM IPTG for 2 hours. Crude lysates of cells expressing lambda gtll recombinants were made as described in Huynh et al. Huynh, T.V. et al., In: DNA Cloning Techniques: A Practical Approach, (D. Glover, ed.) IRL Press, Oxford, Vol. 1, pp. 49-78 20 (1985). Crude lysates of cells expressing plasmid encoded proteins were made by harvesting cells from overnight cultures and resuspending the cells in 10 mM Tris pH7.5/10 mM EDTA containing 100 ug lysozyme/ml. After 10 minutes at room temperature, 25 SDS was added to a final concentration of 0.5%. protease inhibitor (Trasylol, Boehringer Mannheim) was added to all crude lysates at a final concentration of 0.3%. The crude protein preparations 30 were electrophoresed on 10% polyacrylamide-SDS Laemmli gels and the separate proteins electrophoretically transferred to nitrocellulose. Laemmli,
U.K., Nature, 227:680-685 (1970). Towbin, H. et
al., Proceedings of the National Academy of
Sciences, USA, 76:4350-4354 (1979). The immobilized
proteins were reacted with a 1:1000 dilution of
monoclonal antibody IT-13 in TBST overnight at 4°C.
The nitrocellulose filters were then washed, reacted
with peroxidase-conjugated goat anti-mouse immunoglobulin, and developed as described by Niman and
co-workers. Niman, H.L. et al., Proceedings of the
National Academy of Sciences, USA, 80:4949-4953
(1983).

The sequences of 5'-end-labeled restriction fragments of the mycobacterial DNA were determined 15 by a modification of the partial chemical degradation technique of Maxam and Gilbert. M.A.D. et al., Mol. Biol. Evol., 2:1-12 (1985). Maxam, A.M. and W. Gilbert, Proceedings of the National Academy of Sciences, USA, 74:560-564 20 (1976). For the M13/dideoxy sequencing studies, Sau3AI fragments from the mycobacterial DNA inserts were subcloned into the BamHI site of M13mp9. DNA was isolated from the M13 recombinants and subjected to the dideoxy chain termination 25 sequencing reactions. Biggin, M.D. et al., Proceedings of the National Academy of Sciences, <u>USA</u>, <u>80</u>:3963-3965 (1983). Sanger, F. et al., Journal of Molecular Biology, 143:161-178 (1980). The products of the sequencing reactions were electrophoresed on 6% acrylamide/7M urea/0.5-2.5 x

electrophoresed on 6% acrylamide/7M urea/0.5-2.5 \times TBE gradient sequencing gels. The gels were dried

under vacuum and exposed to Kodak XRP-1 film. The nucleotide sequences were determined independently for both strands of the mycobacterial DNA.

Computer-aided analyses of the nucleic acid sequences and deduced protein sequences were 05 performed using the Databases and programs provided by the Nucleic Acid and Protein Identification Resources of the National Institutes of Health as well as the programs of Chou and Fasman and Hopp and Woods. Chou, P.Y. and G.D. Fasman, Adv. Enzym., 10 47:45-148 (1978). Hopp, T.P. and K.P. Woods, Proceedings of the National Academy of Sciences, USA, 78:3824-3828 (1981). The nucleotide sequence of the region containing the M. tuberculosis 65kD gene and the deduced amino acid sequence of the two 15 long open reading frames are represented in Figure 8.

B-galactosidase assays were also carried out. Cells were grown in LB broth or LB broth plus 2.5mM IPTG to an OD₆₀₀ of about 0.3. Crude lysates were made and b-galactosidase activity assayed as described by Miller. Miller, J.H., Experiments in Molecular Genetics, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY (1972).

25 Equivalents

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Those skilled in the art will recognize or be able to ascertain, using no more than routine experimentation, many equivalents to the specific materials and components described herein. Such equivalents are intended to be encompassed in the scope of the following claims.

CLAIMS

- 1. Isolated DNA encoding an immunogenic protein antigen of Mycobacterium tuberculosis.
- 2. DNA of Claim 1 selected from the group consisting of DNA encoding Mycobacterium tuberculosis
 protein antigens of molecular weight 71kD,
 65kD, 19kD, 14kD and 12kD.
- 3. Isolated DNA encoding an immunodominant protein antigen of Mycobacterium tuberculosis, the protein antigen having a molecular weight of approximately 65kD and recognized by a monoclonal antibody selected from the group consisting of: IT-31; C1.1; IIH9; IIC8; T2.3; Y1-2; SA2.D7C and IT-13.
- 15 4. Isolated DNA encoding an immunodominant protein antigen of Mycobacterium tuberculosis, the protein antigen having a molecular weight of approximately 19kD and recognized by a monoclonal antibody selected from the group consisting of: IT-10; IT-12; IT-16; and IT-19.
 - 5. Isolated DNA encoding an immunodominant protein antigen of Mycobacterium tuberculosis, the protein antigen having a molecular weight of approximately 71kD and recognized by the monoclonal antibody IT-11.

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- 6. Isolated DNA encoding an antigenic determinant of Mycobacterium tuberculosis protein.
- 7. DNA of Claim 6 which encodes an antigenic determinant selected from the group consisting of antigenic determinants of Mycobacterium tuberculosis proteins of molecular weight 71kD, 65kD, 19kD, 14kD and 12kD.
- 8. Isolated DNA encoding an amino acid sequence of an antigenic determinant of Mycobacterium
 10 tuberculosis protein, said protein having a molecular weight of approximately 65kD.
 - 9. Isolated Mycobacterium tuberculosis DNA encoding an immunodominant protein antigen having a molecular weight of approximately 65kD, said DNA selected from the group consisting of:
 - a. the DNA insert of clone Y3141;
 - b. the DNA insert of clone Y3143;
 - c. the DNA insert of clone Y3150;
 - d. the DNA insert of clone Y3253; and
- e. the DNA insert of clone Y3262.
 - 10. A protein antigen encoded by DNA of Claim 9.
 - 11. A protein antigen of Claim 10, wherein the protein antigen is recognized by a monoclonal antibody selected from the group consisting of IT-31; Cl.1; IIH9; IIC8; T2.3; Y1-2; SA2.D7C and IT-13.

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- 12. Isolated DNA having a nucleotide sequence selected from the group consisting of: a) the nucleotide sequence represented in Figure 6, or a portion thereof; b) the nucleotide sequence represented in Figure 7, or a portion thereof; and c) the nucleotide sequence represented in Figure 8, or a portion thereof.
- 13. A protein or a peptide selected from the group consisting of: a) proteins or peptides encoded by the nucleotide sequence represented in Figure 6, or a portion thereof; b) proteins or peptides encoded by the nucleotide sequence represented in Figure 7, or a portion thereof; and c) proteins or peptides encoded by the nucleotide sequence represented in Figure 8, or a portion thereof.
 - 14. A peptide having the amino acid sequence of an antigenic determinant of Mycobacterium
 tuberculosis protein, said antigenic determinant being unique to Mycobacterium tuberculosis protein.
 - 15. A peptide encoded by isolated Mycobacterium
 tuberculosis DNA, said peptide recognized by helper T cells.
- 25 16. A peptide encoded by the Mycobacterium tuberculosis DNA insert of clone Y3150 or a portion of said DNA insert.

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- 17. Isolated DNA encoding a protein of Myco-bacterium africanum the protein having a molecular weight of 65kD.
- 18. Isolated DNA encoding a protein of Myco
 bacterium avium, the protein having a molecular weight of 65kD.
 - 19. A vaccine comprising DNA encoding Mycobacterium tuberculosis protein in a recombinant vaccine vector capable of expressing said DNA.
- 10 20. A vaccine of Claim 19 in which the recombinant vaccine vector is vaccinia virus or cultivatable mycobacteria.
- 21. A vaccine of Claim 20 in which the DNA encode's the 65kD Mycobacterium tuberculosis protein recognized by the monoclonal antibody IT-13, or a portion of said protein.
 - 22. A vaccine comprising DNA encoding an antigenic determinant unique to Mycobacterium tubercu-losis cultivatable mycobacteria capable of expressing said DNA.
 - 23. A method of detecting antibody against Myco-bacterium tuberculosis in a biological fluid, comprising the steps of:
- a) incubating an immunoadsorbent comprising a solid phase to which is attached

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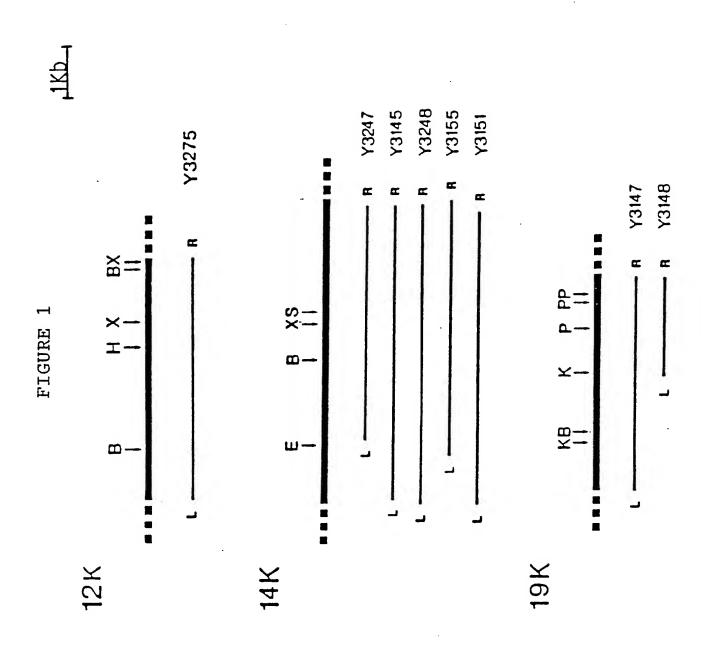
immunodeterminant <u>Mycobacterium tuberculosis</u> protein with a sample of the biological fluid to be tested, under conditions which allow the anti-<u>Mycobacterium tuberculosis</u> antibody in the sample to bind to the immunoadsorbent;

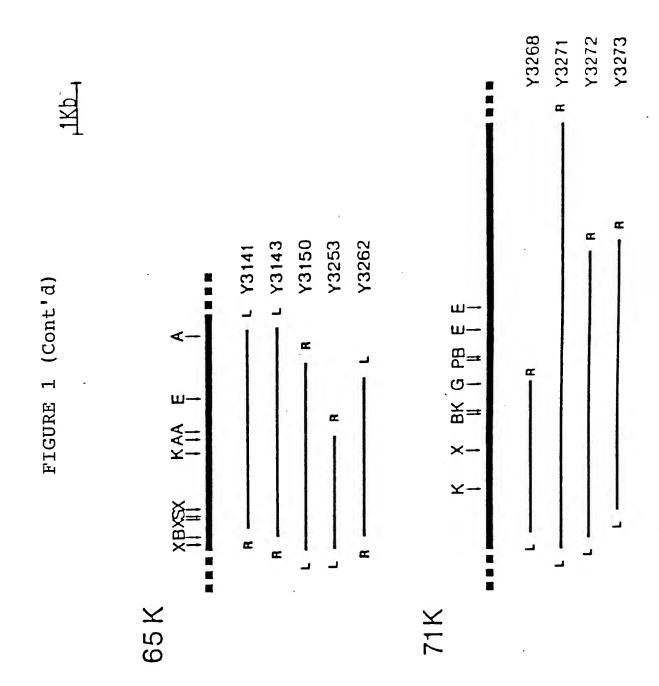
- b) separating the immunoadsorbent from the sample; and
- c) determining if antibody is bound to the immunoadsorbent, as an indication of anti-Mycobacterium tuberculosis in the sample.
- 24. A method of Claim 23 in which the <u>Mycobacterium</u> tuberculosis protein attached to the solid phase has a molecular weight of approximately 65kD.
- 25. A method of detecting antibody against Mycobacterium tuberculosis in a biological fluid, comprising the steps of:
 - a) incubating an immunoadsorbent comprising a solid phase to which is attached a peptide having the amino acid sequence of an antigenic determinant of Mycobacterium
 tuberculosis protein with a sample of the biological fluid to be tested, under conditions which allow antibody against Mycobacterium
 tuberculosis to bind to the immunoadsorbent;
 - b) separating the immunoadsorbent; and
 - c) determining if antibody is bound to the immunoadsorbent, as an indication of the

presence of the antibody against <u>Mycobacterium</u> tuberculosis in the sample.

- 26. A method of Claim 25 in which the peptide has the amino acid sequence of an antigenic determinant which is unique to Mycobacterium tuberculosis protein.
- 27. A kit useful in detecting antibody against

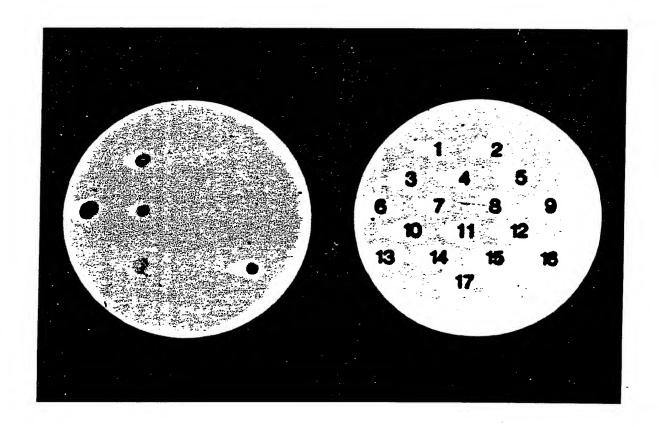
 Mycobacterium tuberculosis in a biological
 fluid, comprising a collection of reagents for
 immunoassay of said antibody, said collection
 of reagents a solid phase to which is attached
 immunodeterminant Mycobacterium tuberculosis
 protein or a peptide having the amino acid
 sequence of an antigenic determinant of
 Mycobacterium tuberculosis.





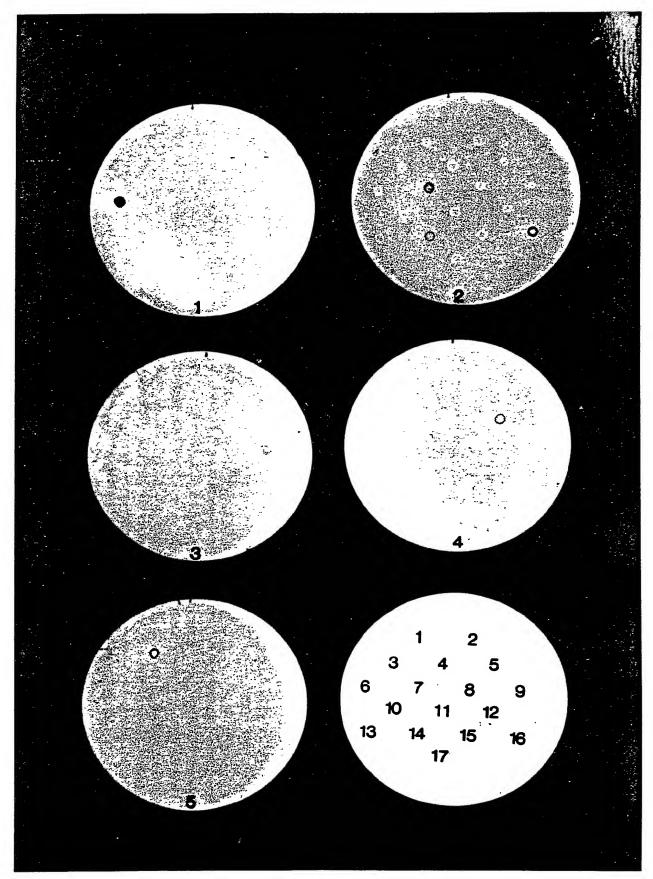
WO 88/05823 PCT/US88/00281

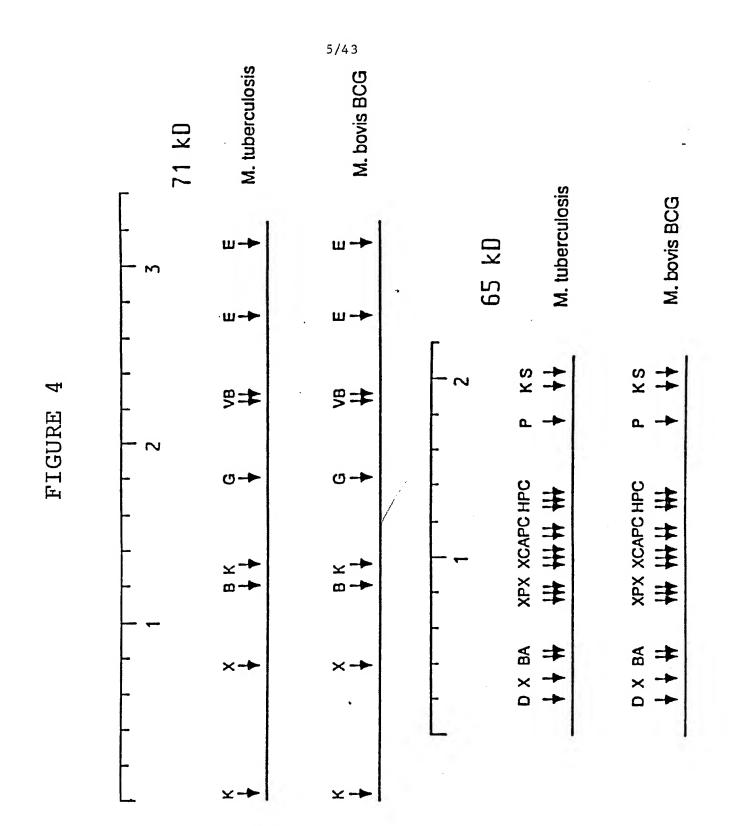
3/43 FIG.2



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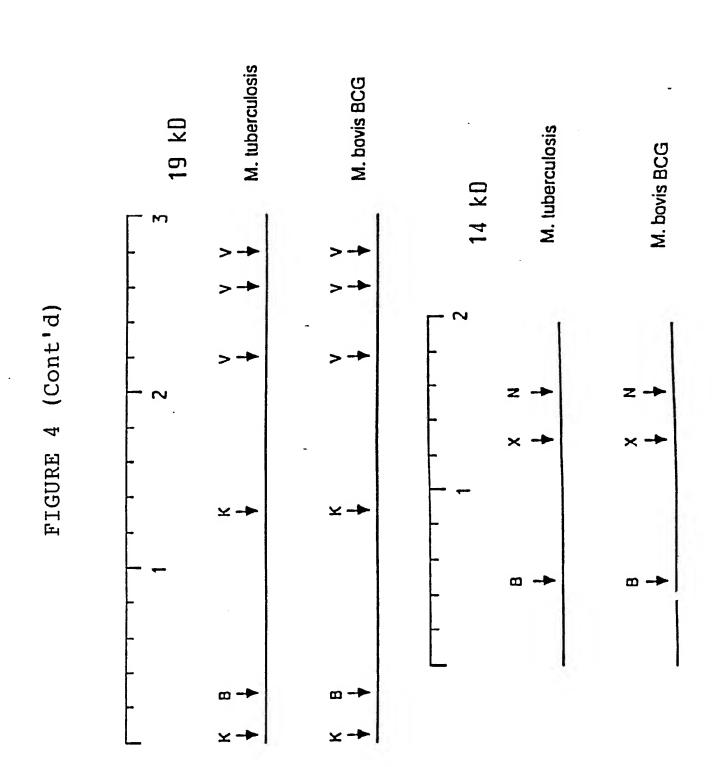
4/43 FIG.3





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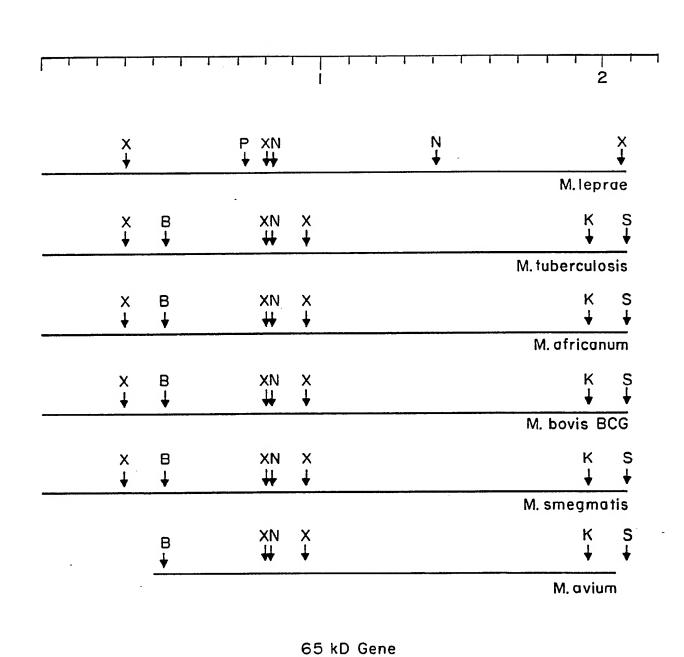


FIG. 5

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FIGURE 6

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CGGCGCGTCCCAGTCCAACACGTCCAATACGCACCGCAAAAGCCGGTACGTGTTGCG Σ Ø 回 Ø 耳 Z Z Н 召 b

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FIGURE 6 (CONT'D)

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FIGURE 6 (CONT'D)

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GCGGTGGAAAGGGCGCAGGTGCAGCCGGACCCACCACTGCGGCTCGTGGGGTGGCTTTAC Ŀı 回 K U U H U 团 α U 460 U U C G Ø C d ტ ഗ C ø Ц

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FIGURE 6 (CONT'D)

TAGCTGTACCGACACCCACATCTACTGGCGCTGGTGCCCCGCCAGCCGAGGCGCCACCCG CGCTATGTTCAGGTCGCGCCGCCGCCGGTGGAGCCACCTGTGGTTGCCCAGCTACTGCTA AACGGGTCGATGACGAI 二 以 召 Ω U 590 S Ø П ш 召 S U 出 ᄓ U 召 K ø, ď Ø S 560 Ø Д Н 耳 Ø Ω ഗ 550 Н 耳 Ø Ω K Н

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(CONT'D) FIGURE

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FIGURE 6 (CONT'D)

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15/43 C 006 TAGCCGCAGCAGCTTTAGGCAACACAGTACGTTGCCATTGCTCACAAGTGGCACACGGCG GACCTACTGCCGTCACCCTCCAAACACAAGGTAGCCGTGATGTAACGGTGATGATGCCAC ATCGGCGTCGTCGAAATCCGTTGTGTCATGCAACGGTAACGAGTGTTCACCGTGTGCCGC Ø 田 耳 H S 只 U S 950 890 3 U H O Z ₽ 880 C, 民 Ø Σ H 3 K U 870 930 Σ 뎨 \mathbf{z} Ω 耳 K CTGGATGACGCCAGTGGGAGGTT Ŀ Ø Z 860 U Ω U Д 耳 口 ĬΉ ഗ Н K Ω U Ø 吖 Ω Ω H 召 Σ ø Ω U 召 Д Д Н Ø

GTGCGGCCATCTACGGCAACCGCTTGGTGCGATGGCTGGTCTTTCTCTCTTAAAAGGCGG GCACCTAGACCTCGGGCCCTGCTAACGCGCATACTGCCGAAGCGGTCCTCAATGCCGATG CGTGGATCTGGAGCCCGGGACGATTGCGCGTATGACGGCTTCGCCAGGAGTTACGGCTAC 耳 \mathbf{z} r 召 耳 Σ Ŀı ᄓ 1010 တ S U α 回 П K K S U K Д Ø 1050 d 民 H 只 ø U വ് ഗ 1040 980 3 Ø CACGCCGGTAGATGCCG1 U 召 U Д 叫 凶 S ഗ U PL, α U Ö 田

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FIGURE

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S CTGGCGATGCTGTCCGTTTCCTCGTGTCCCACTTCGCACCTGACTGCCAGCGCCATCGGC Д GACCGCTACGACAGGCAAAGGAGCACAGGGTGAAGCGTGGACTGACGGTCGCGGTAGCCG K K S Н K H Д K 以 瓦 H щ H ഗ Ø S S Д 出 U H K Ø ഗ Ŀ ഗ U Д K U H K S S Ø U K Д U Н Q U Д 召 Д Ø Н C 回 Ω S 以 Ø K U

CTCGGCGGTAAGACCAGCGTCCAGAAAGGCCTACAAGTTCGTTGTTCAGCTGATGTCCTT 回 Ø 1190 S M, ഗ U Н U Ø Н Z 二 Ω Д 以 Д U ഠ 又 GAGCCGCCATTCTGGTCGCAGGTCT 召 Ω Д Н Ø 1160 凶 S H 3 Д K 回 ĮΉ 1150 \mathbf{z} 3 ρι K Д S Н 回

F

U TCCAGCAGTAGCTGCCATTCCTGGTCTTGCAGTGGCCGAGGCACCACACGTGTTGGCGCC AGGICGICATCGACGGIAAGGACCAGAACGICACCGGCICCGIGGIGIGCACAACCGCGG GCGGTGAGACCACGACCGCGGCAGGCACGACGGCAAGCCCCGGCGCGCGCCTCCGGGCCGA Ø Ü Q C 吖 1250 а 吖 Д H 以 М 以 吖 р 回 ഗ Ċ K Ø Д 召 1290 以 ഗ 召 ഗ U Н H М 召 ĸ ഗ 只 S Z S Ωι 吖 ഗ S H 吖 二 以

TGCCGTTGGGAGGCCTCCACTTCAGGCAACCCGAGCCATTGCAGTTGCCGCAGTGCGACC ACGGCAACCCTCCGGAGGTGAAGTCCGTTGGGCTCGGTAACGTCAACGGCGTCACGCTGG GGCCGTTACAGTTGTAGCGCTAGCCGCCCCCCGCTGGCCGTAACGGCGGCACGAGTGGC ⋈ 以 只 S Ø **1** 1370 民 ы П Ø ഗ Σ C Ø 1360 出 Д ഗ Ľ Δ, 1350 U М ρι K Д ĸ ഗ 1340 ഗ 民 耳 ഗ Σ U H S 出 Σ O Ø 吖 ы

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(CONT'D) 9 FIGURE

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20/43 TCTAGTGACCCTGGCGATGGCCCCAGCTGTACCGGTTGGGCTACAGTGGCCACTTGTTCA CTATGTGCAGCCCGTGGCCTGTCCCATTGCGGAGCCGTTGGTTCCTGCCGTCGGTGATGT CGGCAACCAAGGACGGCAGCCACTACA U ഗ 回 S U C 1490 1550 М ĸ AGATCACTGGGACCGCTACCGGGGTCGACATGGCCAACCCGATGT Ø 1540 U Д U Н Ø EH C Z, 回 GATACACGTCGGCACCGGACAGGGTAACGCCT 以 3 C 1530 Σ N 1470 K 只 H 召 ഗ U U Д G р O O Н ri. 1520 Д 1460 U S 民 K N H Ö 召 Д μ U U ഗ ш 1510 1450 又 Ø Ы Н C S ഗ ĸ Ω

21/43 GCGCAGCCTCGGCCCGTCAGTCCGGATCGCCGCTGCTAAGCTCGCCAACGGTAGGCAG CGCGTCGGAGCCGGGCAGTCAGGCCTAGCGCGGCGACGATTCGAGCGGTTGCCATCCGTC GCAAGCTTTAGCTCCACTGGACAAGGATTGGATTTCGCACAGCTACGCCCGACACTTGTC AAAGCGIGICGAIGCGGGCIGIGAACAG Ø U 团 3 H U Q S Ø Ø U р 吖 Д 召 Н 回 Н 以 召 Ω Þ 1600 Z S S 又 K Z 吖 р M. U U 回 Ø Z > K O 凶 1580 Д Ø ഗ H U K Д K S U 民 ഗ 1570 ഗ U Ŀ 노 义 S 回 H 吖

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GITGIGCGCCTIGACCACAGCGGAGACGATCGCCAGGCCGGAGCCCGGTGCTACCGGCTTG TICACCGITGGCGTGGCGTTTGAGCCATATAGGCCCACTCGATGAGTGCCACTAGCAAGG CAACACGCGGAACTGGTGTCGCCTCTGCTAGCGGTCCGGGCTCGGGCCACGATGGCCGAAC C 回 Z H U A 召 K ĸ U H Н Ω ഗ 二 Ü 1790 ഗ 召 Н H 田 ĸ 回 U ഗ Д H ഗ Н S 回 1780 耳 ш U S Н Ü Д 3 民 U ш U ø 召 ഗ Ω Ω H ഗ Д ps, U Н AAGTGGCAACCGCACCGCAAACT 回 ഗ ഗ Z 1760 1700 S K O Ø 公 二 ⋈ U > Ω 耳 ഗ C Ø K 吖 凶 Н ш 1750 1690 U U Н Ø 田 Z р a Ц Z

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FIGURE 6 (CONT'D)

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23/43 召 U GCCTTATGGGCCGGGTAACAGCTAGTGGACGTCGTGCTGCACGCAGCCGGGGCCACGAGTT CGCCCTGGCACTGCATAGCGGCGCCCGCTTGGCGAGCTTTTGGAGCCTGACGTCGCGCCG Ø S Ø 工 H Н U C 1910 $\boldsymbol{\sigma}$ α Д S K K K H ū 田 1900 以 K K K S u 召 Ø C 1890 1830 K S K > 二 C 出 召 以 1880 U 1820 Ø S Z Ω Σ 3 Ü GCGGGACCGTGACG Ç 召 ഗ 1870 1810 r ĸ ഗ Z C 吖 Д

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FIGURE 6 (CONT'D)

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24/43 GTGGTGGACCACGTTGGCACGGCGTAGTGGGCCCTACTGGTGGCCAAGCCCCCCGTCCAG CGCGCAGCAGTGCTAGCACGGCCCTGGCCACACGTGCGCCCGCAACCGGTCGTCCAACCA GCATCACCCGGGATGACCACCGGTTCGGGGGGGCAGGTC Z U Н М 2030 1970 口 K S K ГJ Ø Z K Ó ¥ 1960 U 吖 Δ S D₄ 耳 2010 1950 二 J EH U ഗ K CACCACCIGGIGCAACCGIGCC K 2000 K 1940 U C 耳 E-1 H Н > 1930 H 吖 工 Н U > 吖 耳

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CTCGCGGGTGACCACTAGACCAGGCCGTTGTACTCGCAGCGACTGGCGCAGTTGGCGTTC A 只 α U > Z GAGCGCCCACTGGTGATCTGGTCCGGCAACATGAGCGTCGCTGACCGCGTCA 2090 召 Ø 召 凶 > Ω O ഗ K 2080 K > d Ø ഗ Σ 耳 \mathbf{Z} 2070 H Z Q Ø U р U ഗ C 3 2060 Ø 区 S S 耳 > C 3 ш Ц Ø ഗ 2050 3 Д G ø C Ø ĸ ഗ ഥ

CCGCGACAIGICCACCGGICCGCGIICCAGCGGCCGCCCCGAGICCAGACGCGCCAGAG ഗ Ø ⊢ Ø ρ S 召 Д C ഗ Ĺτι ഗ 出 Σ r. Д

U 2160 GGCGCTGTACAGGTGGCCAGGCGCAAGGTCGCCGGCGGGGCTCAGGTCTGCGCGGTCGTC 3 U ď 2150 ഗ П 3 U Ω Ц S K r U U K ĸ Ü ш 区 2130 П 3 U 딦 Z H K D 召 Ü 2120 H р 只 U > 3 U 2110 H \mathbf{z} U S 以

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26/43 **⊑**⊣ 2280 CACACGCTGCCGGAGCCCTAGCAGGGGATAGGCGACGCAGTTAAGGCACATTGGTGCCTA GICGICCAGGAGCIGCICIGCACAAIAGGCGAGCCAGAGGCIACGGIGGGCCGAGIAGCG 吖 K 以 Σ H S 回 U S K U Z K р 召 Д G U Д Ü > 出 凶 C t 回 Ø C 二 \mathbf{Z} O 以 S 2200 四 U S ᄓ Н Ø Н K 口 2190 d 以 Д C 凶 U U Z Ω GTGTGCGACGGCCTCGGGATCGT CAGCAGGICCICGACGAGACGI 召 E-I Ω α 2180 ഗ ഗ Н 回 以 召 K H 口 Д S D₁ G 召 S Ø 回 U 口 2230 召 U K Ω Н 只 Ø Н U Ø H 二 Ø >

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FIGURE 6 (CONT'D)

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27/43 AAGTGACCGGACGGCGCGGCTGTCACGCCGTCGCTACACCAGCTCGTAGGCCAACTCGCG GCGGCGTTCCCCTCAAGCGTCAAGTACTGACCGTAGCCGTTGCTTGACCGCGTGTGCCA TTCACTGGCCTGCCGCGCCGACAGTGCGGCAGCGATGTGGTCGAGCATCCGGTTGAGCGC CGCCGCAAGGGGAGTTCGCAGTTCATGACTGGCATCGGCAACGAACTGGCGCACACGGGT 召 只 回 S Ø Z Д 2330 2390 召 U ഗ \mathbf{Z} Ø 出 K ഗ 口 回 Ŀı S K 2380 2320 K 召 Σ ഗ ø Ø ഗ 2310 O Ø ഗ K 出 二 E-I Н Ø 回 C 2360 ഗ 又 C 吖 N Z 召 召 H C ഗ Ø М 2290 2350 U Д Ø 口 O K ഗ ы Ŀı

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FIGURE 6 (CONT'D)

2460 TCGGGCACCCGGACCGG GCTGGGCTCAACGGGGTGGAGCCACCTCCCCAAACGCAGTCCAAGCCCGTGGGCCTGGCC > G S 召 Д ď Д U ш ы 出 2450 C U Q, 召 Д K S [L] Ŀı Z CGACCCGAGTTGCCCCACCTCGGTGGAGGGGTTTGCGTCAGGT ĸ D. 2440 Н E--1 K Ω Ø Z H 2430 U Д C 딥 U S K Z S H 召 ഥ 2420 ĸ U K E > 耳 3 Ü Д U Д G Ø 口 2410 Z EH 口 ഥ Ω 区 Д ø U Д Ü ഗ × K

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Ø 2510 K K 回 ഗ 24 回 2500 ഗ S S K 2490 Д Н Д Ц U 2480 ĸ ĸ U 回 2470 × S

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FIGURE 6 (CONT'D)

GCGGCGTTGCCGCGGTCGCGGGCCGAACTACTACTGCTAAGGCCGCCAGCAGCAGCGCCGCTGG TGATGATGACGATTCCGGCGGTCGTCGCGGCGACC G α Д 以 d K 2570 Ω Ω U H р K U K Д K 2560 U ഥ Z ഗ K H ഗ \mathbf{z} 耳 2550 出 Σ ഗ Ø Ы ഗ Ø or, Д 2540 U Ø S Q 3 U CGCCGCAACGGCGC K Ø r 2530 K 召 > Z K ø, N

2640 <u>AACGCAA:TCACCGTGACGATTCCGAAAATGATCAGCATCTGCAACATCGTGGCGTCGACG</u> TIGCGITAGIGGCACIGCTAAGGCITITACTAGICGTAGACGITGTAGCACCGCAGCIGC പ്പ 以 二 2630 吖 Ω K 2620 Ω Ø ഗ S Ω S 2610 耳 Γı S Д S 团 Z 2600 ഗ 区 H 耳 α K U 2590 Ω ĸ Z

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FIGURE 7

			3	0/43			
48		96	,	144		192	
GAG	Glu	ATC	Ile	ATC	Ile	299	G1y
		299	G1y	GAC	Asp		
GAG	Glu	ACC	Thr	TTC	Phe	CGC	G1y
GGG	G1y	CTG	Leu	ACT	Thr	AAG	Lys
GTG CAG ATC CAG GTC TAT CAG GGG GAG CGT	Val Gln Ile Gln Val Tyr Gln Gly Glu Arg	GAG CTG ACC GGC	Lys Leu Leu Gly Ser Phe Glu Leu Thr Gly Ile	GAG GTC ACT TTC GAC ATC 144	Ile Glu Val Thr Phe Asp	GTG CAC GTC ACC GCC AAG GAC AAG GGC ACC	Thr Ala Lys Asp Lys Gly Thr
TAT	TYr	TTC	Phe	GAG	Glu	AAG	Lys
GIC	Val	GGG TCC TTC	Ser	ATC	Ile	CCC	Ala
CAG	Gln	999	G1Y	CAG	Gln	ACC	\mathtt{Thr}
ATC	Ile	CTC	Leu	SCG	Pro	GTC	Val
CAG	Gln	AAG TTG CTC	Leu	GGG ATT CCG CAG ATC	Gly Ile Pro Gln	CAC	Val His Val
GTG	Val	AAG	Lys	999	G1y	GTG	Val
TCG	Ser	AAC	Asn	550	Arg	ATT	Ile
CCG	Pro	CAC	His	S S S S S S S S S S S S S S S S S S S	Pro	GGC	Gly Ile
TTC CAA	Gln	9 29	Ala	909 80	Ala	GCC AAC	Asp Ala Asn
	Phe	CCC	Ala	SCG	Pro	GCC	Ala
GAA	G1n	49 ATC GCC	Ile	CCG	Pro	GAC	Asp
М		49		97		145	

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FIGURE 7 (CONT'D)

240		. 288	51,715	336		384	
GAA	Glu	ATC AAG GAC GCC GAA GCG CAC GCC GAG GAG GA	Asp	TTG	Leu	CCT	Gly
AAG	$ ext{Lys}$	GAG	Glu	ACA	Thr	GAG	Glu
TCC	Ser	GAG	Glu	GAG	Glu	GCC	Ala
·CTG	Leu	GCC	Ala	CCC	Ala	GAG	Glu
CGA ATC CAG GAA GGC TCG GGC CTG TCC AAG GAA	Arg Ile Gln Glu Gly Ser Gly Leu Ser Lys	CAC	Ile Lys Asp Ala Glu Ala His Ala Glu Glu Asp	GAG GCC GAT GTT CGT AAT CAA GCC GAG ACA	Glu Ala Asp Val Arg Asn Gln Ala Glu	AAG TTC GTC AAA GAA CAG CGT GAG GCC GAG	Lys Phe Val Lys Glu Gln Arg Glu Ala Glu Gly
TCG	Ser	909	Ala	AAT	Asn	CAG	Gln
CCC	Gly	GAA	Glu	CGT	Arg	GAA	Glu
GAA	Glu	ರಿದಿ	Ala	GTT	Val	AAA	Lys
CAG	Gln	GAC	Asp	GAT	Asp	GTC	Val
ATC	Ile	AAG	Lys	೦೦೮	Ala	TTC	Phe
_	Arg			GAG	Glu	AAG	Lys
ATC	II e	ATG	MET	GAG	Glu	GAG	Glu
ACG	Thr	CGC	Arg	CGC	Arg	ACG	Thr
GAG AAC ACG	Glu Asn	GAC	Asp	CGT	Arg	CAG	Tyr Gln
GAG		GAC ATT	Ile	AAG	Lys	TAC	
AAG	Lys	GAC	Asp	CGC	Arg	GTC	Val
193		241		289		337	

FIGURE 7 (CONT'D)

432	32, 0 8 7	/43 & C LC	576	
GTG Val	CAA Gln	AGC Ser	CCA	
GCG Ala	CAT His	GCA Ala	TGC Cys	ιν
GCC Ala	GGC	$\tt GGG$	CGC	61
GAT Asp	TTC	TCT Ser	TGG	GGC G1y
GTT Val	TAT Tyr	$ ext{GGC}$	CAC	CTC
AAG Lys	GGA Gly	GCA Ala	GGC	CGG
AAC Asn	ATC Ile	GTC Val	ACA Thr	CCC
CTG	CGG	GGA GTC Gly Val	GTC Val	CCA
ACG Thr	TGG	CCA Pro	TGC	TGC
GAC Asp	ACT Thr	$_{\rm GGG}$	GGC	CGG
GAA Glu	GGC	GCT Ala	TCA	$\tt GGG$
CCT	$ ext{GGC}$	GAA Glu	AGC	GCC Ala
GTA Val	GAA Glu	GGA	AGC	CGA Arg
AAG Lys	GCG Ala	GGC GAT Gly Asp	CGA Arg	CGG CGG CGA Arg Arg Arg
TCG Ser	GAA Glu	GGC Gly	CTA Leu	CGG
GGT	GCG Ala	GTC Val	GAT Asp	CCC
3 8 5	433	481	529	577

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FIGURE 8

TCGAACGAGGGGCGTGACCCGGTGCGGGGCTTCTTGCACTCGGCATAGGCGAGTGCTAAG AGCTTGCTCCCCGCACTGGGCCACGCCCCGAAGAACGTGAGCCGTATCCGCTCACGATTC 10 50 60	AATAACGTTGGCACTCGCGACCGGTGAGTGCTAGGTCGGGACGGTGAGGCCAGGCCCGTC TTATTGCAACCGTGAGCGCTGGCCACTCACGATCCAGCCCTGCCACTCCGGTCCGGGCAG 100 110 120	GTCGCAGCGAGTGGCAGCGAGGACAACTTGAGCCGTCCGT	AGCGTAAGTAGCGGGGTTGCCGTCACCCGGTGACCCCGGTTTCATCCCGGATCGGGAGGA TCGCATTCATCGCCCCAACGGCAGTGGGCCACTGGGGGCAAAGTAGGGGCTAGGCCTCCT 190 200 210 220	N H F A M A K T I A Y D E E A R R G L E ATCACTTCGCAATGGCCAAGACAATTGCGTACGACGAGGGCCCGTCGCGGCCTCGAGC TAGTGAAGCGTTACCGGTTCTTAACGCATGCTGCTTCTCCGGGCAGCGCCGGAGCTCG 250 250 270 280 300	R G L N A L A D A V K V T L G P K G R N GGGGCTTGAACGCCCTCGCCGATGCGGTAAAGGTGACATTGGGCCCCAAGGGCCGCAACG CCCCGAACTTGCGGGAGCGGCTACGCCATTTCCACTGTAACCCGGGGTTCCCGGCGTTGC 310 320 330 340 350	V V L E K K W G A P T I T N D G V S I A TCGTCCTGGAAAAGAAGTGGGGTGCCCCACGATCACCAACGATGGTGTCCATCGCCA AGCAGGACCTTTTCTTCACCCCACGGGGGGTGCTAGTGGTTGCTACCACACAGGTAGCGGT 370 380 390 400 410 420

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>	AGGAGATCGAGCTGGAGGATCCGTACGAGAAGATCGGCGCCGAGCTGGTCAAAGAGGTAG	TCCTCTAGCTCGACCTCCTAGGCATGCTCTTCTAGCCGCGGCTCGACCAGTTTCTCCATC	480
ш	CAC	010	
ᅩ	AAA		
>	GIC	CAG	470
_	CTG	GAC	
ш	GAG	CIC	
∢)))	550	480
5	ესე	9))	ব
;	ATC	TAG	
¥	AAG	TIC	0
ш	GAG	CIC	450
> -	TAC	ATG	
ے	500	299	
٥	GAT	CTA	440
ш	GAG	CIC	
ب	CTG	GAC	
Ш	GAG	CTC	430
H	ATC	TAG	4
ш	GAG	CTC	
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FIGURE

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GCAACCAAGCGCTCCCGGACGCGTTGCAGCGCCGGCGCGGGTTGGGCGAGCCAGAGTTTG 800 CGTTGGTTCGCGAGGGCCTGCGCAACGTCGCGGCCGCCCAACCCGCTCGGTCTCAAAC ے م ¥ 0 580 G

.GCTCAAGGGCGCCAAGGAGG CGCCGTAGCTTTTCCGGCACCTCTTCCAGTGGCTCTGGGACGAGTTCCCGGGGTTCCTCC GCGGCATCGAAAAGGCCGTGGAGAAGGTCACCGAGACCC¯

V E T K E Q I A A T A A I S A G D Q S I TCGAGACCAAGGAGCAGTTGCGGCCACCGCAGCGATTTCGGCGGGTGACCAGTCCATCG <u> AGCTCTGGTTCCTCGTCTAACGCCGGTGGCGTCGCTAAAGCCGCCCACTGGTCAGGTAGC</u> G D L I A E A M D K V G N E G V I T V E GTGACCTGATCGCGAGGCGATGGACAAGGTGGCAACGAGGGCGTCATCACCGTCGAGG CACTGGACTAGCGGCTCCGCTACCTGTTCCACCCGTTGCTCCCGCAGTAGTGGCAGTCC Ţ

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TCAGGTTGTGGAAACCCGACGTCGAGCTCGAGTGGCTCCCATACGCCAAGCTGTTCCCGA AGTCCAACACCTTTGGGCTGCAGCTCGAGCTCACCGAGGGTATGCGGTTCGACAAGGGC G 810 800 G

TGTAGAGCCCCATGAAGCACTGGCTGGGCCTCGCAGTCCTCCGCCAGGACCTCCTGGGGA CCTGGAGGACCCC **ACATCTCGGGGTACTTCGTGACCGACCCGGAGCGTCAGGAGGCGG**1 880 ш 0 880

960 | CCACTGTCAAGGATCTGCTGCCGCTGCTCGAGA TGTAGGACGACCAGTCGAGGTTCCACAGGTGACAGTTCCTAGACGACGGCGACGAGGTCT ۵ S S

TCCAGTAGCCTCGGCCATTCGGCGACGACTAGTAGCGGCTCCTGCAGCTCCCGCGCTCCGCG FCGCCGAGGACGTCGAGGGCGAGGCGC G **AGGTCATCGGAGCCGGTAAGCCGCTGCTGATCA**T ۵. G G

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080 ACAGGTGGGACCAGCAGTTGTTCTAGGCGCCGTGGAAGTTCAGCCACCGCCAGTTCCGAG FCGTCAACAAGATCCGCGGCACCTTCAAGTCGGTGGCGGTCAAGGCTC 1060 ~

GGCCGAAGCCGCTGGCGGCGTTCCGCTACGACGTCCTATACCGGTAAGAGTGGCCACCAG TCGGCGACCGCCGCAAGGCGATGCTGCAGGATATGGCCA Œ 0 J C

TGCGGCTGGACAGCGACGATCCGT

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G	<u>GG1</u>	CCA	
∢	225	000	_
G	2001	000	1250
ш	GAG	CTC	
>)T5	3CAC	
—	CATO	JTA(1240
-	CAC	3TG(
-	SACC	TG(
ш	CGAC	3010	30
0	3GA(CTO	1230
×	CAA(TT(
—	CAC	3TG	80
>	3GT(CA(1220
>	100	3CA(•
> >	3GT(CCA	
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FIGURE

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T D A I A G R V A Q I R Q E I E N S D S CCGACGCCATCGCCGGACGAGTGGCCCAGGAGATCGAGAACAGCGACTCCG GGCTGCGGTAGCGGCCTGCTCACCGGGTCTAGGCGGTCCTCTAGCTCTTGTCGCTGAGGC 1290 1280 1270

D Y D R E K L Q E R L A K L A G G V A V ACTACGACCGTGAGAGCTGCAGGAGCTGGCCAAGCTGGCCGGTGGTGTGCGGGTGA TGATGCTGGCACTCTTCGACGTCCTCGCCGACCGGTTCGACCGGCCACCACACGCCACT 1370 1360 1350 1340

1440 TCAAGGCCGGTGCCGCCACCGAGGTCGAACTCAAGGAGCGCAAGCACCGCATCGAGGATG <u> AGTTCCGGCCACGGCGGTGGCTCCAGCTTGAGTTCCTCGCGTTCGTGGCGTAGCTCCTAC</u> 1430 エ œ 1420 1410 1400 G 1390

1500 GCCAAGCGTTACGGTTCCGGCGGCAGCTCCTCCCGTAGCAGCGGCCACCCCCCACACTGCG CGGTTCGCAATGCCAAGGCCGCCGTCGAGGAGGGCATCGTCGCCGGTGGGGGTGTGACG(1490 G 1480 ш 1460

1580 1550 TGTI

CCAACAT

FIGURE 8 (CONT'D)

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1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	CGA ACT 740	A CCG CGC BØØ	G GTG CAC BBB	1928 1928	CTT GAA 98Ø	CGA GCT Ø4Ø
GAA CTT 16	V GGT CCA(E AGG	1000	366 300 1	GCAACCTT CGTTGGAA 1986	GTA CAT
L ICT IGA	CAAC	CGA GCT	CAT GTA	TTGTGGC	20 150	(7 ()
H G L N ACGGACTGAACG TGCCTGACTTGC	V GGTC CCAC	T 25 2	GACT	TT	CATCO 178	GATG CTAC
	$\alpha \cup C \alpha$	T TGG' 798	GCGCCACATGGG CGCCGCTGTACCC 1850	CCCGGTCCCTTT GGGCCAGGGAAA		0000 0000 0000
700	ACCO TGG	("()	6 0 0 0 0 0 0 1 8	STC CAG	TCG AGC	200 000 000
A L P A G ACCTGCCGGCTGG TGGACGCCGACC	TGCTG/	F L TTCCTC	V P G G GTTCCCGGTGG CAAGGGCCACC 1840	000	A G	AGC(TCG(
000	TG	CA	ا ددد 1900 1900	3 2 2 2 3 2 3 3 3 3 3 3 3 3 3 3 3 3 3 3	ACGC TGCG	₹
9 000 000 660	G V CGGCGTTC CCGCAAC	CGA CGA 782	TC(AG(846	CGAGGAGCC SCTCCTCGC 1900	000	GCAG(CGTC(2020
CTCT	6 2 2 2 2 1	2000	CGT 3CA	GAG CTC	CAC GTG	
N AAC TTG	TGCC ACGG	A G L F CGCGGGGCTGTT GCGCCCGACAA	S TC(9,0	GAACAC CTTGTC) ()
V R N GGTGCGCAACC CCACGCGTTGG 1850	A CCT CGA		A S V SGCTTCCGTTC SCGAAGGCAAG	GCA CGT	3	T C A G
V TGC ACC 856	D L L A SATCTGCTCGCT CTAGACGAGCGA	S TCCA AGGT/ 1770	Q Q (i)	T C A G B 9	00 00 00 05	
1000	rgc Acg	2 CA	E K	GAAG	GTA CAT	CCG GGC 2
K GAAG CTTC	TC]	N A A A A A A A A A A A A A A A A A A A		CGGCGAGA GCCGCTCT BBØ	000	$\mathcal{G}_{\mathcal{G}}$
CGAG/	GGA CCT Ø	TGC ACG	AAA TTT	000	CTACO GATGO Ø	000
A GCC CGG 84Ø	CGA(CGA)	- I	E K E SGAAAAGGA SCTTTTCC1 1820	200 200 188	GAGCT CTCGA 1940	TC AG ØØ
V V A CGTGGTGGCC GCACCGG	\ _	CAG GTC	K P. AGCCGC	1881 1880 1881	(70)	CT
V T G C A C (V TCT AGA	L TG AC	K AAG TTC	CTGA(STTG	ნეე ენნ
	GTG	A CGC GGG GGG GGG GGG GGG GGG GGG GGG GG	D AC TG	F TCJ AGA	TGG ACC	T G A C
P (CGGCGCGCGCGGCGGCGGCGGGGGGGGGGGGGGGGGG	T CCG GGC 169(GG CC 75	CC CC B1	TT A A 8 7	CTC GAG 193	CTG GAC 199
р ОСС ССС 1	GAC CTG	S TTC AAG	TGC ACG	GGA CCT	TC(AG(666 1
GA C.T	CAC	R CG CG	CA CA	AT TA	<u> </u>	99
L TG AC	CT	T 000	V TC AG	9)))	TG

FIGURE 8 (CONT'D)

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CACCGCAGCGGGCGGTGTCGTCATCGGGGCCTGCGTCCGACGCCTGGGCACGGCCGTCGA GTGGCGTCGCCCGCCACAGCAGTAGCCCCGGACGCAGGTGCGGACCCGTGCGGCAGCT 2050 2050 2060 2060 2000 2000	CGATCAGCGAGTAGCCGCTAGGATCGGATGGCGGCCACAACAGGGTGACTTCGCTGCGGT GCTAGTCGCTCATCGCGATCCTAGCCTACCGCCGGTGTTGTCCCACTGAAGCGACGCCA 2110 2120 2130 2130 2140 2150	GGGCCAGGTTTTGCCGCGTACGACCCCCGATCAGGCCGACGTCGACCACTGCCCGGGGTC CCCGGTCCAAAACGGCGCATGCTGGGGGCTAGTCCGGGCTGCAGCTGGTGACGGGCCCCAG 2170 2180 2190 2200	CATCGGGGCCGTCGGGGAGTTCGCGCACCGGCTCGACTGCCACCGTGTGCACGCGAT GTAGCCCCGGCAGCCCTTCAAGCGCGTCGTGGCCGAGCTGACGGTGGCACACGTGCGCTA 2230 2240 2250 2260	2390 2340 2310 2320 2330 2340 6GCCATCATCGACGGTAGTCGGGCAAGGCGGCCAGCC CGGTAGTCGGGTAGTCGGGCAAGGCGGCGGCCAGCC	2350 2350 2370 2380 2390 2400 CAAACTCCACGGATGTGTTTACTC CAAACTCCACGGTGTGTTACTC CAAAACTCCACGGGTGTGTGTTACTC CAAAACTCCACGTGGGTGCCTAAGCTCCTATCCGCGGGCTACACAATGAG	2480 2480 2480 2480 2480 2480 2480 2480	70 2480 2490 251 TCGACTTGGGGATGCCGGGGGGGTACTCGGCTCACGG	Y F N S K P I G S G P I S P E R V A I N
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CAAGGTCTTCGACCACAACTCCGACGGACGCGACGGCTCCGGGCGCAACTAACAGGGGCT

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2580

2540

(CONT'D)

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FIGURE

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7	40 11 10 10 10 10 10 10 10 10 10 10 10 10	ØØ AG TC L	39/43 89 Y 50 S	128 164 167 5	388 566 366 A
	2848 2848 2848 2848 2848 2848 2848 2848	2880 2870 2680 2700 AGGCCGCCGCGCGCGCGCGCGCGCGCGCGCGCGCGCGCG	AGGACGTTCGCCAGGCCGTGTTGGAGACCGCCCGTTGATCCGAGGGCGGA TCCTGCAAGCGGTCCGGCACTCTGGCGGGCAACTAGGCTCCCGCCT TCTGCAAGCGGTCCGGCACTCTGGCGGGCAACTAGGCTCCCGCCT	2920 2980 2910 2820 CCCGAACTCAAAGCCGCCGTGCTCATGCCGCCGGTGCGTAGCCGGCGGCGGCGGCGGCGGCGGCGGCGGCGGCGGCGGC	2840 2850 2880 2870 2880 2870 2880 CCGCCTCCGAGCCAGCCGCGCTTCCTAAGGCGGCGTTTTGCATCCCCGCGCGCG
	Ø GGGT CCCA P N	GTGT GACA T N	G L	STAGG	78 TTGC/ AACGI
Ŋ	263 FACCG \TGGC	289 26660 30006	27E FTGAT AACTA S	28] TGGC(ACCG(CGTT CCATT GCAA
≻	CCGCT GGCGA G	GCGAC CGCTC A V	1 8 (2005) (2005)	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	A A A
S	2820 ACTG TGAC	268Ø 366CG 3CCGC	274Ø 3ACCG .TGGC	2800 TGCCG ACGGC	2888 CTAAG 3ATTC
· · · · · · ·	3 CGGC	GCCGC CGGCC A P	TGGAC ACCTC	CTCA' GAGT	CTTC GAAG(S G
Z	818 AGCCI TCGG	678 ACTGO	738 GTGT CACA,	790 CGTG	2850 2000 3000 A
	A G C C	SGAGC CCTCG	2 AGGCC FCCGG	2 3 5 5 6 6 7 8	CCAGGGGTCG
S	30000 30000 30000 6 8	BBACAGITGE	28 CGCCA GCGG1 A L	BØ CAAA(GTTT(40 CGAG GCTC
Z	286 TTGC(AACG(286 (2000)	27: CGTT GCAA	27 SAACT TTGA	28 GCCTC CGGAG A E
S D N S G S N V F S P Y S T G P S	GGGT SCCCA P N	AGGC TCCG	SAGGA STCCT L V	S 5 00000 00000	800
ם פ ר	2598 166666 166666	2858 GAGTCC CTCAGG S D	2710 AGTTGATCTCAAC	277 GGAT CCTA	283 CCAA GGTT
2000 R L	TCTAAC AGATTC R L	GCCTGA CGGACT G S	s 9	GGCGA(CCGCT(A L	GCTGA CGACT S V

40/43

9 0 0 0 0 0 0 0	36
2950 2980 2970 2980 2990 36 GGTCCCGATGCCGCTGTTCAGGGAGCCCGAATTCCCGATGCCGATGTTTCCGCTGCC CCAGGGCTACGGCGACAAGTCCCTCGGGCTTAAGGGCTACGGCTACAAAGGCGACGG T G I G S N L S G S N G I G I N G S G	050
299Ø TTTCC AAAGG N G	3050
2S TGTT ACA/	36
CGA GCT,	
298Ø GATGC CTACG I G	3040
29 CGA GCT	36
TTC(AAG(! !
297Ø CCGAA GGCTT S	3010 3020 3030 3040 3
AGCCCG TCGGGC	36
GGA CCT S	
2980 TTCAG AAGTC N L	3020
28 TGT1 ACAA	36
	•
295Ø GATGC CTACG I G	3010
25 CCGA 3GCT	36
GTC(CAG(T	j
00	;

(CONT'D

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FIGURE

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BBB GGA CCT S 080 GTTGAATAAGCCGACGTTGCCGGTGCCCGAGTTCCCGAAGCCGATGTTGCCGCTACCCGA CAACTTATTCGGCTGCAACGGCCACGGGCTCAAGGGCTTCGGCTACAACGGCGATGGGCT z G J Z S G J Z

3120 CAACTTCGGCGGCTTTGGGTAGACCACTAGTGGCCACTAGGGCTTGGGCTATAAGGGCGA GTTGAAGCCGCCGAAACCCATCTGGTGATCACCGGTGATCCCGAACCCGATATTCCCGCT 3110 G J 3090 × 3080 G G 3180 TGGCCACAACGGCTTCGGCTATAAGGGCAGCGGCTCCAACGGCTCCGGGTCCAACGGCGA TGCCGCT **ACCGGTGTTGCCGAAGCCGATATTCCCGTCGCCGAGGTTGCCGAGGCCCAGGT** 3170 J 3160 3150 J z 3140 z

CGGCCACAACGGCGACGGCTACAACGGCCACGGCCACAACGGCGACGGCTACAACAA <u> GCCGGTGTTGCCGCTGCCGATGTTGCCGGTGCCGGTGTTGCCGCTGCCGATGTTGTTGT</u> Z 3220 z 3210 Z 3200 3190 z

TGCCGAA CGGCTACAACAACAAGGGCTACAACAACAAGGGCTACAAGGGCGAGGGCCACAAGGGCTT GCCGATGTTGTTGCCGATGTTGTTGTTGCCGATGTTGCCGCTGCCGGT 3290 3280 z 3330 3270 z z z 3260 Z Z 3310 3250 z

3360 CGGGTCTAACTAGACCGGCAAGAACGGCTACAGCTACGGCTCCAAGGCGTTCTGGACGAC 3350 GCCCAGATIGATCTGGCCGTTCTTGCCGATGTCGATGCCGAGGT 3340 9 G 8 (CONT'D)

FIGURE

41/43

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	CCAGGGCGCCAGTTGTGCGACGGCCGCAGACGCATCGAAGTGGTAACCAGCCATCGCCGC	GGTCCCGCGGTCAACACGCTGCCGGCGTCTGCGTAGCTTCACCATTGGTCGGTAGCGGCG	WPALQAVAASADFHYGAMAA		CACGTCCAATGCCCACATTTGCTCGTATGCCGCCTCGACGTCCATGAGCGCCGGAGCGTT	GTGCAGGTTACGGGTGTAAACGAGCATACGGCGGAGCTGCAGGTACTCGCGGCCTCGCAA	V D L A W M Q E Y A A E V D M L A P A N		CTGCCCAAACCAGTTCGTAGCTGCCAGCAGCTGCATCAGGCCACGATTGGCCGCTACCAC	3AC	QGFWNTAALLGRNAAVV		TGCCGGCTGCACGGTGGCCGCCAGCGCCGCCTCGAACGCGGTCGCTGTTGCCATGGCCTG	<u> ACGGCCGACGTGCCACCGGCGGTCGCGGCGGAGCTTGCGCCAGCGACAACGGTACCGGAC</u>	∢	
	C	9			J	9				_			,	~		

3780 CTGCCTCGGTTCGCTGCGATAACTGCGCTCGTTAAGAAGCCGGTCGAGCGGGGTCCGCCA V S G L S A I S A L L E E A L E G W A T CTGCCGGTAGTAGCGGCGGCGCCTGCCTGGGTCGGTCCGCGGTGATCAGTCAAGCCTACA TCTTCGGCCAGCTCGCCCCAGGCGGT GACGGAGCCAAGCGACGCTAT Z

A C G C C G G C G A A A G G C G G C G G C G G C C C G G T C C G A T C C A T G A C C C A A C G

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42/43

FIGURE 8 (CONT'D)

3840	CTC	GAG	ш
ന	GAT	CTA	H
	ATT	TAA	z
3830	CGA	GCT	S
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	DYD.	GTC	_
0	CAT	GTA	×
3820	AAA	TTT	L.
	CCC	000	⋖
	ACC	TGG	IJ
3810	CGG	000	<u>م</u>
ന	CCC	ggg	IJ
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3800	1CC	AGG	IJ
38	550	225	ط
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3790	AGC	TCG	∀
		505	~
	GGCCGCAGCAATTAGCGGTCCCGACCCGGGACCGGCAAACATCAGTGCCGAATTGATCTC	CCGGCGTCGTTAATCGCCAGGGCTGGGCCTTGGCGTTTGTAGTCACGGCTTAACTAGAG	∢

GGCGGCAACCACGCAAAATGCGGGCTTGTCAGCCGATCCAACTTAACTGTCAGCGACCG エ

<u> AACGGCACCGCCATAGCCGTGAAGTTATGGTGAGTAGAAACCCCAGTAGAAACCTCGCGG</u> <u> TGCCGTGGCGGTATCGGCACTTCAATACCACTCATCTTTGGGGTCATCTTTGGAGCGCC</u> 3950 3940 ≥ 3930 3920 > 3910 ۵.

CCTAGGAACCGCCAGCTTACCTAGTCCCGGGTAGGGGCCGACTGGCGGCCGGGATGCAGC <u>~</u> 4010 م م 4000 G م 3990 ن۵ 3980 3970

<u> ACTCCCAGACGGTGGACGGGGCATTACAGCGACCATACCGTTCGTGGCTGCGGCGCCGGG</u> TGAGGGTCTGCCACCTGCCCCGTAATGTCGCTGGTATGGCAAGCACCGACGCCGCGGGCCC 4070 2 4060 م ď 4050 ~ G 4040 J 4030 8

AAGAGTTGCTCCGCGACGCGTTCACCCGGTTGATCGAACATGTCGACGAACTCACCGACG TTCTCAACGAGGCGCTGCGCAAGTGGGCCAACTAGCTTGTACAGCTGCTTGAGTGGCTGC 4110 4100 œ 4090

GCCTCACCGACCAACTCGCCTGCTACCGCCCGACCCCCAGCGCCAACAGCATTGCGTGGC CGGAGTGGCTGGTTGAGCGGACGATGGCGGGCTGGGGGTCGCGGTTGTCGTAACGCACCG 4160 4160

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FIGURE 8 (CONT'D)

GCACAGCGCCCGGGTGCAGGATATACAGGTCGCCCATGTGGCCGGCGTG GCACAGCGCCCACGTCCTATATGTCCAGCGGGTACACCGGCCGCAC CGTGTCGCGGGCCCACGTCCTATATGTCCAGCGGGTACACCGGCCGCAC 270 4280 4290 4300 4310 GACCCGCGACGGTTGGGTGGACCGCTTTGGGTTAGATCTGCCGCGGCAC	CCGTGCTGT	TAGACGGCG 4370 CACCCGCCG	CTGGGCGCTGCCAACCCACCTGGCGAAACCCAATCTAGACGGCGCCGTGCTGT 330 4340 4350 4360 4360 4370 TGGACACCGTCCCGAGGATGTGGCGAAGGTACGGGAACCACGCGAAGGATTC	CCACCTGGCO 4350 AGGATGTGGC	GCGCTGCCAAC 434Ø CACCGTCCCGA	TCCACACCTGGG 4330 CCGGATATGGA	
TGCTCTGGCACAGCGCCCGGGTGCAGGATATACAGGTCGCCCATGTGGCCGGCGTGGAAG ACGAGACCGTGTCGCGGGCCCACGTCCTATATGTCCAGCGGGTACACCGGCCGCGCTTTC	432Ø GGCACGACA CCGTGCTGT	431Ø ATCTGCCGC TAGACGGCG	43ØØ CTTTGGGTTAG, SAAACCCAATC	4290 GGTGGACCG	428Ø CGCGACGGTTC GCGCTGCCAAC	AGTGTG CCACAC	
0107 BYOY BOOK BOOK	428Ø GCGTGGAAG GCACCTTC	4250 ATGTGGCCG TACACCGGC	424Ø ACAGGTCGCCC, TGTCCAGCGGG	423Ø FGCAGGATAT, ACGTCCTATA	4220 AGCGCCCGGG TCGCGGGCCC/	4210 TGCTCTGGCAC, ACGAGACCGTG	